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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁶: A61K 38/18, 39/395, 38/17, 35/12 // C07K 14/475, 16/22, C12N 15/12, C12P 21/08	A2	(11) International Publication Number: WO 99/39729 (43) International Publication Date: 12 August 1999 (12.08.99)
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(54) Title: USE OF HEREGULIN AS AN EPITHELIAL CELL GROWTH FACTOR (57) Abstract Ligands which bind to the HER2, HER3 and/or HER4 receptors are useful as normal epithelial cell growth factors.		

USE OF HEREGULIN AS AN EPITHELIAL CELL GROWTH FACTOR

BACKGROUND OF THE INVENTION**Field of the Invention**

5 This invention relates to the use of HER2, HER3 and/or HER4 ligands, in particular heregulin polypeptides, as epithelial cell growth factors.

Description of Background and Related Art

10 The HER (ErbB) family belongs to the subclass I receptor tyrosine kinase superfamily and consists of three distinct receptors, HER2, HER3, and HER4. A ligand for this ErbB family is the protein heregulin (HRG), a multidomain containing protein with at least 15 distinct isoforms.

15 Transduction of signals that regulate cell growth and differentiation is regulated in part by phosphorylation of various cellular proteins. Protein tyrosine kinases are enzymes that catalyze this process. Receptor protein tyrosine kinases are believed to direct cellular growth via ligand-stimulated tyrosine phosphorylation of intracellular substrates. Growth factor receptor protein tyrosine kinases of the class I subfamily include the 170 kDa epidermal growth factor receptor (EGFR) encoded by the *erbB1* gene. *erbB1* has been causally implicated in human malignancy. In particular, increased expression of this gene has been observed in more aggressive carcinomas of the breast, bladder, lung and stomach.

20 The second member of the class I subfamily, p185^{neu} was originally identified as the product of the transforming gene from neuroblastomas of chemically treated rats. The *neu* gene (also called *erbB2* and HER2) encodes a 185 kDa receptor protein tyrosine kinase. Amplification and/or overexpression of the human HER2 gene correlates with a poor prognosis in breast and ovarian cancers (Slamon *et al.*, *Science* 235:177-182 (1987); and Slamon *et al.*, *Science* 244:707-712 (1989)). Overexpression of HER2 has been correlated with other carcinomas including carcinomas of the stomach, endometrium, salivary gland, lung, 25 kidney, colon and bladder. Accordingly, Slamon *et al.* in US Pat No. 4,968,603 describe and claim various diagnostic assays for determining HER2 gene amplification or expression in tumor cells. Slamon *et al.* discovered that the presence of multiple gene copies of HER2 oncogene in tumor cells indicates that the disease is more likely to spread beyond the primary tumor site, and that the disease may therefore require more aggressive treatment than might otherwise be indicated by other diagnostic factors. Slamon *et al.* conclude that the HER2 gene amplification test, together with the determination of lymph node status, 30 provides greatly improved prognostic utility.

A further related gene, called *erbB3* or HER3, has also been described. See US Pat. No. 5,183,884; Kraus *et al.*, *Proc. Natl. Acad. Sci. USA* 86:9193-9197 (1989); EP Pat Appln No 444,961A1; and Kraus *et al.*, *Proc. Natl. Acad. Sci. USA* 90:2900-2904 (1993). Kraus *et al.* (1989) discovered that markedly elevated 35 levels of *erbB3* mRNA were present in certain human mammary tumor cell lines indicating that *erbB3*, like *erbB1* and *erbB2*, may play a role in human malignancies. Also, Kraus *et al.* (1993) showed that EGF-dependent activation of the ErbB3 catalytic domain of a chimeric EGFR/ErbB3 receptor resulted in a proliferative response in transfected NIH-3T3 cells. This is now believed to be the result of endogenous ErbB1 or ErbB2 in NIH-3T3. Furthermore, these researchers demonstrated that some human mammary tumor cell lines display a significant elevation of steady-state ErbB3 tyrosine phosphorylation further 40 indicating that this receptor may play a role in human malignancies. The role of *erbB3* in cancer has been explored by others. It has been found to be overexpressed in breast (Lemoine *et al.*, *Br. J. Cancer* 66:1116-1121 (1992)), gastrointestinal (Poller *et al.*, *J. Pathol.* 168:275-280 (1992), Rajkumar *et al.*, *J. Pathol.* 170:271-278 (1993), and Sanidas *et al.*, *Int. J. Cancer* 54:935-940 (1993)), and pancreatic cancers (Lemoine *et al.*, *J. Pathol.* 168:269-273 (1992), and Friess *et al.*, *Clinical Cancer Research* 1:1413-1420 (1995)). 45

The class I subfamily of growth factor receptor protein tyrosine kinases has been further extended to

do not have the complete glycosylated spacer region between the Ig-like domain and EGF-like domain. Only one of the GGFs, GGFII, possessed a N-terminal signal peptide. See also WO 92/18627; WO 94/00140; WO 94/04560; WO 94/26298; and WO 95/32724 which refer to GGFs and uses thereof.

Ho *et al.* in *J. Biol. Chem.* 270(4):14523-14532 (1995) describe another member of the heregulin family called sensory and motor neuron-derived factor (SMDF). This protein has an EGF-like domain characteristic of all other heregulin polypeptides but a distinct N-terminal domain. The major structural difference between SMDF and the other heregulin polypeptides is the lack in SMDF of the Ig-like domain and the "glyco" spacer characteristic of all the other heregulin polypeptides. Another feature of SMDF is the presence of two stretches of hydrophobic amino acids near the N-terminus.

While the heregulin polypeptides were first identified based on their ability to activate the HER2 receptor (see Holmes *et al.*, *supra*), it was discovered that certain ovarian cells expressing *neu* and *neu*-transfected fibroblasts did not bind or crosslink to NDF, nor did they respond to NDF to undergo tyrosine phosphorylation (Peles *et al.*, *EMBO J.* 12:961-971 (1993)). This indicated another cellular component was necessary for conferring full heregulin responsiveness. Carraway *et al.* subsequently demonstrated that ¹²⁵I-

rHRGβ1₁₇₇₋₂₄₄ bound to NIH-3T3 fibroblasts stably transfected with bovine *erbB3* but not to non-transfected parental cells. Accordingly, they conclude that ErbB3 is a receptor for HRG and mediates phosphorylation of intrinsic tyrosine residues as well as phosphorylation of ErbB2 receptor in cells which express both receptors. Carraway *et al.*, *J. Biol. Chem.* 269(19):14303-14306 (1994). Sliwkowski *et al.*, *J. Biol. Chem.* 269(20):14661-14665 (1994) found that cells transfected with HER3 alone show low affinities for heregulin, whereas cells transfected with both HER2 and HER3 show higher affinities.

This observation correlates with the "receptor cross-talking" described previously by Kokai *et al.*, *Cell* 58:287-292 (1989); Stern *et al.*, *EMBO J.* 7:995-1001 (1988); and King *et al.*, 4:13-18 (1989). These researchers found that binding of EGF to the EGFR resulted in activation of the EGFR kinase domain and cross-phosphorylation of p185^{HER2}. This is believed to be a result of ligand-induced receptor heterodimerization and the concomitant cross-phosphorylation of the receptors within the heterodimer (Wada *et al.*, *Cell* 61:1339-1347 (1990)).

Plowman and his colleagues have similarly studied p185^{HER4}/p185^{HER2} activation. They expressed p185^{HER2} alone, p185^{HER4} alone, or the two receptors together in human T lymphocytes and demonstrated that heregulin is capable of stimulating tyrosine phosphorylation of p185^{HER4}, but could only stimulate p185^{HER2} phosphorylation in cells expressing both receptors. Plowman *et al.*, *Nature* 336:473-475 (1993).

The biological role of heregulin has been investigated by several groups. For example, Falls *et al.*, (discussed above) found that ARIA plays a role in myotube differentiation, namely affecting the synthesis and concentration of neurotransmitter receptors in the postsynaptic muscle cells of motor neurons. Corfas and Fischbach demonstrated that ARIA also increases the number of sodium channels in chick muscle. Corfas and Fischbach, *J. Neuroscience*, 13(5): 2118-2125 (1993). It has also been shown that GGFII is mitogenic for subconfluent quiescent human myoblasts and that differentiation of clonal human myoblasts in the continuous presence of GGFII results in greater numbers of myotubes after six days of differentiation (Sklar *et al.*, *J. Cell Biochem.*, Abst. W462, 18D, 540 (1994)). See also WO 94/26298 published November 24, 1994.

Holmes *et al.*, *supra*, found that HRG exerted a mitogenic effect on mammary cell lines (such as SK-BR-3 and MCF-7). The mitogenic activity of GGFs on Schwann cells has also been reported. See, e.g., Brockes *et al.*, *J. Biol. Chem.* 255(18):8374-8377 (1980); Lemke and Brockes, *J. Neurosci.* 4:75-83 (1984); Lemke and Brockes, *J. Neurosci.* 4:75-83 (1984); Brockes *et al.*, *Ann. Neurol.* 20(3):317-322 (1986); Brockes, J., *Methods in Enzym.*, 147: 217-225 (1987) and Marchionni *et al.*, *supra*. Schwann cells constitute

87:6378-6382 (1990). However, early efforts to select high affinity fusion phage failed, presumably due to the polyvalence of the phage particles. This problem was solved with the development of a "monovalent" phage display system in which the fusion protein is expressed at a low level from a phagemid and a helper phage provides a large excess of wild-type coat protein. Bass *et al.*, *Proteins* 8:309-314 (1990); Lowman *et al.*, *Biochem.* 30:10832-10838 (1991). Monovalent phage display can be used to generate and screen a large number of variant polypeptides to isolate those that bind with high affinity to a target of interest.

Approximately 50,000 infants are born in the United States every year with birth weights, less than 1.5 kg. About two thirds of these very low birth weight infants have evidence of pulmonary immaturity manifested as respiratory distress shortly after birth. The majority of these infants require mechanical ventilation. Respiratory distress syndrome, caused by insufficient pulmonary surfactant production, as well as structural immaturity of the lung, is responsible for respiratory difficulties observed in these prematurely born neonates. Well developed alveoli are necessary to provide efficient oxygen transfer from the air-liquid interface of the lung to the systemic circulation. Surfactant proteins are critical in reducing the alveolar surface tension at low lung volumes and preventing alveolar collapse.

A need continues to exist for a method of treatment for respiratory distress syndrome and other diseases associated with immature lung development and low lung surfactant production.

SUMMARY OF THE INVENTION

In general an object of the invention is to provide a method of inducing epithelial cell growth and development for the purpose of promoting repair and healing of tissue damage or injury.

Accordingly, one object of this invention is to provide a method of treating respiratory distress syndrome in patients, primarily human patients, in need of such treatment. A further object is to provide a method of inducing lung epithelial cell growth and development. A further object is to provide a method of increasing lung surfactant protein A production in the lung of persons with impaired oxygen transfer in the lung alveoli. This invention is useful in treating infants/neonates with respiratory distress as well as youth and adults with poor lung function due to lung injury or damage.

In one aspect of this invention, it has now been discovered that these objects and the broader objective of treating conditions associated with epithelial cell damage and injury are achieved by administering to a patient in need of such treatment an effective amount of a heregulin ligand, preferably a polypeptide or fragment thereof. These heregulin (HRG) polypeptides, include HRG- α , HRG- β 1, HRG- β 2, HRG- β 3 and other HRG polypeptides which cross-react with antibodies directed against these family members and/or which are substantially homologous as defined below and includes HRG variants such as N-terminal and C-terminal fragments thereof. A preferred HRG is the ligand disclosed in Fig. 1A - 1D and further designated HRG- α . Other preferred HRGs are the ligands disclosed in Figure 2A - 2E, and designated HRG- β 1; disclosed in Figure 3A - 3E designated HRG- β 2; and disclosed in Figure 4A - 4C designated HRG- β 3.

In another aspect, the invention provides a method in which HRG agonist antibodies are administered to achieve the objects of the invention. In this embodiment, HER2/HER3 or fragments thereof (which also may be synthesized by *in vitro* methods) are fused (by recombinant expression or an *in vitro* peptidyl bond) to an immunogenic polypeptide and this fusion polypeptide, in turn, is used to raise antibodies against a HER2/HER3 epitope. Agonist antibodies are recovered from the serum of immunized animals. Alternatively, monoclonal antibodies are prepared from *in vitro* cells or *in vivo* immunized animals in conventional fashion. If desired, the agonist antibodies may be obtained by phage display selection from a phage library of antibodies or antibody fragments. Preferred antibodies identified by routine screening will bind to the receptor, but will not substantially cross-react with any other known ligands such as EGF, and will activate the HER receptors HER2, HER3 and/or HER4. In addition, antibodies may be selected that are

ID NO:12) of γ -HRG obtained as described in U.S. Serial No. 08/891,845. The hydrophobic region is underlined. The EGF-like domain is shaded, cysteine residues in the EGF-like domain are circled. N-linked glycosylation sites are marked above the nucleic acid sequence with a ().

Figure 8 shows the cDNA sequence (SEQ ID NO:13) and amino acid sequence (SEQ ID NO:14) of SMDF obtained as described in U.S. Serial No. 08/339,517. An EGF-like domain and the apolar and uncharged domains (*i.e.* "apolar I" consisting of residues from about 48-62 and "apolar II" consisting of residues from about 76-100) are underlined. Cysteines in the EGF-like domain and in the "cysteine knot" in the unique N-terminal domain ("NTD-cys knot") are boxed. The stop codon is denoted by the letter "O".

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

HRG ligands, in particular polypeptides and agonist antibodies thereof, have affinity for and stimulate the HER2, HER3 and/or HER4 receptors or combinations thereof in autophosphorylation. Included within the definition of HRG ligands, in addition to HRG- α , HRG- β 1, HRG- β 2, HRG- β 3 and HRG- β 2-like, are other polypeptides binding to the HER2, HER3 and/or HER4 receptor, which bear substantial amino acid sequence homology to HRG- α or HRG- β 1. Such additional polypeptides fall within the definition of HRG as a family of polypeptide ligands that bind to the HER2, HER3 and/or HER4 receptors.

Heregulin polypeptides bind with varying affinities to the HER2, HER3 and/or HER4 receptors. Generally, the HER3 and HER4 receptors are bound with high affinity. It is also known that heterodimerization of HER2 with HER3 and of HER2 with HER4 occurs with subsequent receptor cross-phosphorylation as described above. In the present invention, epithelial cell growth and/or proliferation is induced when a heregulin protein interacts and binds with an individual receptor molecule or a receptor dimer such that receptor phosphorylation is induced. Binding and activation of HER2, HER3, HER4 or combinations thereof, therefore, is meant to include activation of any form of the receptor necessary for receptor activation and biologic function including monomeric receptor and dimeric receptor forms. Dimeric receptor forms may be referred to below, for example, as HER2/HER3, HER2/HER4, and HER3/HER4.

I. DEFINITIONS

In general, the following words or phrases have the indicated definition when used in the description, examples, and claims.

"Heregulin" (HRG) ligand is defined herein to be any isolated ligand, preferably a polypeptide sequence which possesses a biological property of a naturally occurring HRG polypeptide. Ligands within the scope of this invention include the NDF, ARIA and GGF growth factor heregulin proteins identified above as well as the SMDF and HRG polypeptides discussed in detail herein. These isolated NDF, ARIA and GGF heregulin polypeptides are well known in the art. HRG includes the polypeptides shown in Figs. 1A-1D, 2A-2E, 3A-3E, 4A-4C, 5A-5D, 6A-6C, 7A-7C and 8 and mammalian analogues thereof. Included are HRG variants such as the γ -HRG described in WO 98/02541, published 22 January 1998; the variants described in WO 98/35036, published 13 August 1998; and the SMDF variants described in US Patent NO. 5,770,567 granted 23 June 1998 (WO 96/15244). These applications are incorporated herein in their entirety. These variants can be prepared by the methods described below, optionally together with alanine scanning and phage display techniques known in the art. Cunningham and Wells, Science 244:1081-85 (1989); Bass et al., Proteins 8:309-14 (1990); Lowman et al., Biochem. 30:10832-38 (1991).

The term a "normal" epithelial cell means an epithelial cell which is not transformed, *i.e.*, is non-cancerous and/or non-immortalized. Further, the normal epithelial cell is preferably not aneuploid. Aneuploidy exists when the nucleus of a cell does not contain an exact multiple of the haploid number of chromosomes, one or more chromosomes being present in greater or lesser number than the rest. Typical properties of transformed cells which fall outside the scope of this invention include the ability to form

in its native conformation generally is HRG as found in nature which has not been denatured by chaotropic agents, heat or other treatment that substantially modifies the three dimensional structure of HRG as determined, for example, by migration on nonreducing, nondenaturing sizing gels. Antibody used in this determination may be rabbit polyclonal antibody raised by formulating native HRG from a non-rabbit species in Freund's complete adjuvant, subcutaneously injecting the formulation, and boosting the immune response by intraperitoneal injection of the formulation until the titer of anti-HRG antibody plateaus.

Ordinarily, biologically or antigenically active HRG will have an amino acid sequence having at least 75% amino acid sequence identity with a given HRG sequence, more preferably at least 80%, even more preferably at least 90%, and most preferably at least 95%. Identity or homology with respect to an HRG sequence is defined herein as the percentage of amino acid residues in the candidate sequence that are identical with HRG residues in the HRG of Fig. 6A-6C, after aligning the sequences and introducing gaps, if necessary, to achieve the maximum percent homology, and not considering any conservative substitutions as part of the sequence identity. None of N-terminal, C-terminal or internal extensions, deletions, or insertions into HRG sequence shall be construed as affecting homology.

Thus, the biologically active and antigenically active HRG polypeptides that are the subject of this invention include each entire HRG sequence; fragments thereof having a consecutive sequence of at least 5, 10, 15, 20, 25, 30 or 40 amino acid residues from HRG sequence; amino acid sequence variants of HRG sequence wherein an amino acid residue has been inserted N- or C-terminal to, or within, HRG sequence or its fragment as defined above; amino acid sequence variants of HRG sequence or its fragment as defined above has been substituted by another residue. HRG polypeptides include those containing predetermined mutations by, e.g., site-directed or PCR mutagenesis, and other animal species of HRG polypeptides such as rabbit, rat, porcine, non-human primate, equine, murine, and ovine HRG and alleles or other naturally occurring variants of the foregoing and human sequences; derivatives of HRG or its fragments as defined above wherein HRG or its fragments have been covalently modified by substitution, chemical, enzymatic, or other appropriate means with a moiety other than a naturally occurring amino acid (for example a detectable moiety such as an enzyme or radioisotope); glycosylation variants of HRG (insertion of a glycosylation site or deletion of any glycosylation site by deletion, insertion or substitution of appropriate amino acid); and soluble forms of HRG, such as HRG-GFD or those that lack a functional transmembrane domain.

"Isolated" means a ligand, such as HRG, which has been identified and separated and/or recovered from a component of its natural environment. Contaminant components of its natural environment are materials which would interfere with diagnostic or therapeutic uses for HRG, and may include proteins, hormones, and other substances. In preferred embodiments, HRG will be purified (1) to greater than 95% by weight of protein as determined by the Lowry method or other validated protein determination method, and most preferably more than 99% by weight, (2) to a degree sufficient to obtain at least 15 residues of N-terminal or internal amino acid sequence by use of the best commercially available amino acid sequenator marketed on the filing date hereof, or (3) to homogeneity by SDS-PAGE using Coomassie blue or, preferably, silver stain. Isolated HRG includes HRG in situ within recombinant cells since at least one component of HRG natural environment will not be present. Isolated HRG includes HRG from one species in a recombinant cell culture of another species since HRG in such circumstances will be devoid of source polypeptides. Ordinarily, however, isolated HRG will be prepared by at least one purification step.

In accordance with this invention, HRG nucleic acid is RNA or DNA containing greater than ten bases that encodes a biologically or antigenically active HRG, is complementary to nucleic acid sequence encoding such HRG, or hybridizes to nucleic acid sequence encoding such HRG and remains stably bound to it under stringent conditions.

Preferably, HRG nucleic acid encodes a polypeptide sharing at least 75% sequence identity, more preferably at least 80%, still more preferably at least 85%, even more preferably at 90%, and most preferably

"Restriction enzyme digestion" of DNA refers to catalytic cleavage of the DNA with an enzyme that acts only at certain locations in the DNA. Such enzymes are called restriction endonucleases, and the sites for which each is specific is called a restriction site. The various restriction enzymes used herein are commercially available and their reaction conditions, cofactors, and other requirements as established by the enzyme suppliers are used. Restriction enzymes commonly are designated by abbreviations composed of a capital letter followed by other letters representing the microorganism from which each restriction enzyme originally was obtained, and then a number designating the particular enzyme. In general, about 1 mg of plasmid or DNA fragment is used with about 1-2 units of enzyme in about 20 ml of buffer solution. Appropriate buffers and substrate amounts for particular restriction enzymes are specified by the manufacturer. Incubation of about 1 hour at 37°C is ordinarily used, but may vary in accordance with the supplier's instructions. After incubation, protein or polypeptide is removed by extraction with phenol and chloroform, and the digested nucleic acid is recovered from the aqueous fraction by precipitation with ethanol. Digestion with a restriction enzyme may be followed with bacterial alkaline phosphatase hydrolysis of the terminal 5' phosphates to prevent the two restriction cleaved ends of a DNA fragment from "circularizing" or forming a closed loop that would impede insertion of another DNA fragment at the restriction site. Unless otherwise stated, digestion of plasmids is not followed by 5' terminal dephosphorylation. Procedures and reagents for dephosphorylation are conventional as described in sections 1.56-1.61 of Sambrook *et al.*, (*Molecular Cloning: A Laboratory Manual* New York: Cold Spring Harbor Laboratory Press, 1989).

"Recovery" or "isolation" of a given fragment of DNA from a restriction digest means separation of the digest on polyacrylamide or agarose gel by electrophoresis, identification of the fragment of interest by comparison of its mobility versus that of marker DNA fragments of known molecular weight, removal of the gel section containing the desired fragment, and separation of the gel from DNA. This procedure is known generally. For example, see Lawn *et al.*, *Nucleic Acids Res.* 9:6103-6114 (1981), and Goeddel *et al.*, *Nucleic Acids Res.* 8:4057 (1980).

"Northern analysis" is a method used to identify RNA sequences that hybridize to a known probe such as an oligonucleotide, DNA fragment, cDNA or fragment thereof, or RNA fragment. The probe is labeled with a radioisotope such as ³²P, or by biotinylation, or with an enzyme. The RNA to be analyzed is usually electrophoretically separated on an agarose or polyacrylamide gel, transferred to nitrocellulose, nylon, or other suitable membrane, and hybridized with the probe, using standard techniques well known in the art such as those described in sections 7.39-7.52 of Sambrook *et al.*, *supra*.

"Ligation" refers to the process of forming phosphodiester bonds between two nucleic acid fragments. To ligate the DNA fragments together, the ends of the DNA fragments must be compatible with each other. In some cases, the ends will be directly compatible after endonuclease digestion. However, it may be necessary to first convert the staggered ends commonly produced after endonuclease digestion to blunt ends to make them compatible for ligation. To blunt the ends, the DNA is treated in a suitable buffer for at least 15 minutes at 15°C with about 10 units of the Klenow fragment of DNA polymerase I or T4 DNA polymerase in the presence of the four deoxyribonucleotide triphosphates. The DNA is then purified by phenol-chloroform extraction and ethanol precipitation. The DNA fragments that are to be ligated together are put in solution in about equimolar amounts. The solution will also contain ATP, ligase buffer, and a ligase such as T4 DNA ligase at about 10 units per 0.5 mg of DNA. If the DNA is to be ligated into a vector, the vector is first linearized by digestion with the appropriate restriction endonuclease(s). The linearized fragment is then treated with bacterial alkaline phosphatase, or calf intestinal phosphatase to prevent self-ligation during the ligation step.

"Preparation" of DNA from cells means isolating the plasmid DNA from a culture of the host cells. Commonly used methods for DNA preparation are the large and small-scale plasmid preparations described

"Protein microsequencing" was accomplished based upon the following procedures. Proteins from the final HPLC step were either sequenced directly by automated Edman degradation with a model 470A Applied Biosystems gas phase sequencer equipped with a 120A PTH amino acid analyzer or sequenced after digestion with various chemicals or enzymes. PTH amino acids were integrated using the CHROMPERFECT data system (Justice Innovations, Palo Alto, CA). Sequence interpretation was performed on a VAX 11/785 Digital Equipment Corporation computer as described (Henzel *et al.*, *J. Chromatography* 404:41-52 (1987)). In some cases, aliquots of the HPLC fractions were electrophoresed on 5-20% SDS polyacrylamide gels, electrotransferred to a PVDF membrane (PROBLOTT, ABI, Foster City, CA) and stained with Coomassie Brilliant Blue (Matsudaira, P., *J. Biol. Chem.* 262:10035-10038, 1987). The specific protein was excised from the blot for N terminal sequencing. To determine internal protein sequences, HPLC fractions were dried under vacuum (SPEEDVAC), resuspended in appropriate buffers, and digested with cyanogen bromide, the lysine-specific enzyme Lys-C (Wako Chemicals, Richmond, VA) or Asp-N (Boehringer Mannheim, Indianapolis, Ind.). After digestion, the resultant peptides were sequenced as a mixture or were resolved by HPLC on a C4 column developed with a propanol gradient in 0.1% TFA before sequencing as described above.

"Antibodies" (Abs) and "immunoglobulins" (Igs) are glycoproteins having the same structural characteristics. While antibodies exhibit binding specificity to a specific antigen, immunoglobulins include both antibodies and other antibody-like molecules which lack antigen specificity. Polypeptides of the latter kind are, for example, produced at low levels by the lymph system and at increased levels by myelomas.

Papain digestion of antibodies produces two identical antigen-binding fragments, called "Fab" fragments, each with a single antigen-binding site, and a residual "Fc" fragment, whose name reflects its ability to crystallize readily. Pepsin treatment yields an $F(ab')_2$ fragment that has two antigen-combining sites and is still capable of cross-linking antigen.

"Fv" is the minimum antibody fragment which contains a complete antigen-recognition and -binding site. This region consists of a dimer of one heavy chain and one light chain variable domain in tight, non-covalent association. It is in this configuration that the three hypervariable regions of each variable domain interact to define an antigen-binding site on the surface of the V_H - V_L dimer. Collectively, the six hypervariable regions confer antigen-binding specificity to the antibody. However, even a single variable domain (or half of an Fv comprising only three hypervariable regions specific for an antigen) has the ability to recognize and bind antigen, although at a lower affinity than the entire binding site.

The Fab fragment also contains the constant domain of the light chain and the first constant domain (CH1) of the heavy chain. Fab' fragments differ from Fab fragments by the addition of a few residues at the carboxyl terminus of the heavy chain CH1 domain including one or more cysteine(s) from the antibody hinge region. Fab'-SH is the designation herein for Fab' in which the cysteine residue(s) of the constant domains bear a free thiol group. $F(ab')_2$ antibody fragments originally were produced as pairs of Fab' fragments which have hinge cysteines between them. Other chemical couplings of antibody fragments are also known.

The "light chains" of antibodies (immunoglobulins) from any vertebrate species can be assigned to one of two clearly distinct types, called kappa (κ) and lambda (λ), based on the amino acid sequences of their constant domains.

Depending on the amino acid sequence of the constant domain of their heavy chains, immunoglobulins can be assigned to different classes. There are five major classes of immunoglobulins: IgA, IgD, IgE, IgG, and IgM, and several of these may be further divided into subclasses (isotypes), e.g., IgG1, IgG2, IgG3, IgG4, IgA, and IgA2. The heavy-chain constant domains that correspond to the different classes of immunoglobulins are called α , δ , ϵ , γ , and μ , respectively. The subunit structures and three-dimensional

antigen binding. For a review of sFv see Pluckthun in *The Pharmacology of Monoclonal Antibodies*, vol. 113, Rosenberg and Moore eds. Springer-Verlag, New York, pp. 269-315 (1994).

The term "diabodies" refers to small antibody fragments with two antigen-binding sites, which fragments comprise a heavy chain variable domain (V_H) connected to a light chain variable domain (V_L) in the same polypeptide chain ($V_H - V_L$). By using a linker that is too short to allow pairing between the two domains on the same chain, the domains are forced to pair with the complementary domains of another chain and create two antigen-binding sites. Diabodies are described more fully in, for example, EP 404,097; WO 93/11161; and Hollinger *et al.*, *Proc. Natl. Acad. Sci. USA* 90:6444-6448 (1993).

The expression "linear antibodies" when used throughout this application refers to the antibodies described in Zapata *et al. Protein Eng.* 8(10):1057-1062 (1995). Briefly, these antibodies comprise a pair of tandem Fd segments ($V_H - C_H1 - V_H - C_H1$) which form a pair of antigen binding regions. Linear antibodies can be bispecific or monospecific.

II. USE AND PREPARATION OF HRG SEQUENCES

H. PREPARATION OF HRG SEQUENCES, INCLUDING VARIANTS

The system to be employed in preparing HRG sequence will depend upon the particular HRG sequence selected. If the sequence is sufficiently small HRG may be prepared by *in vitro* polypeptide synthetic methods. Most commonly, however, HRG will be prepared in recombinant cell culture using the host-vector systems described below. Suitable HRG includes any biologically active and antigenetically active HRG.

In general, mammalian host cells will be employed, and such hosts may or may not contain post-translational systems for processing HRG preprosequences in the normal fashion. If the host cells contain such systems then it will be possible to recover natural subdomain fragments such as HRG-GFD from the cultures. If not, then the proper processing can be accomplished by transforming the hosts with the required enzyme(s) or by supplying them in an *in vitro* method. However, it is not necessary to transform cells with the complete prepro or structural genes for a selected HRG when it is desired to only produce fragments of HRG sequences such as an HRG-GFD. For example, a start codon is ligated to the 5' end of DNA encoding an HRG-GFD, this DNA is used to transform host cells and the product expressed directly as the Met N-terminal form (if desired, the extraneous Met may be removed *in vitro* or by endogenous N-terminal demethionylases). Alternatively, HRG-GFD is expressed as a fusion with a signal sequence recognized by the host cell, which will process and secrete the mature HRG-GFD as is further described below. Amino acid sequence variants of native HRG-GFD sequences are produced in the same way.

HRG sequences located between the first N-terminal mature residue and the first N-terminal residue of HRG-GFD sequence, termed HRG-NTD, may function at least in part as an unconventional signal sequence or as a normally circulating carrier/precursor for HRG-GFD having unique biological activity.

HRG-NTD is produced in the same fashion as the full length molecule but from expression of DNA in which a stop codon is located at the C-terminus of HRG-NTD. In addition, HRG variants are expressed from DNA encoding protein in which both the GFD and NTD domains are in their proper orientation but which contain an amino acid insertion, deletion or substitution at the GFD-NTD cleavage site (located within the sequence VKC) which inhibits or prevents proteolytic cleavage of the NTD-GFD joining site *in vivo*, and wherein a stop codon is positioned at the 3' end of the GFD- encoding sequence. In an example of this group of variants (termed HRG-NTDXGFD), (1) the lysine residue found in the NTD-GFD joining sequence VKC is deleted or (preferably) substituted by another residue other than arginyl such as histidyl, alanyl, threonyl or seryl and (2) a stop codon is introduced in the sequence RCT or RCQ in place of cysteinyl, or threonyl (for HRG- α) or glutaminy (for HRG- β).

screening genomic DNA libraries include, but are not limited to, oligonucleotides; cDNAs or fragments thereof that encode the same or a similar gene; and/or homologous genomic DNAs or fragments thereof. Screening the cDNA or genomic library with the selected probe may be conducted using standard procedures as described in chapters 10-12 of Sambrook *et al.*, *supra*.

5 An alternative means to isolate the gene encoding HRG- α is to use polymerase chain reaction (PCR) methodology as described in section 14 of Sambrook *et al.*, *supra*. This method requires the use of oligonucleotide probes that will hybridize to HRG- α . Strategies for selection of oligonucleotides are described below.

10 Another alternative method for obtaining the gene of interest is to chemically synthesize it using one of the methods described in Engels *et al.* (*Agnew. Chem. Int. Ed. Engl.*, 28: 716-734, 1989). These methods include triester, phosphite, phosphoramidite and H-Phosphonate methods, PCR and other autoprimer methods, and oligonucleotide syntheses on solid supports. These methods may be used if the entire nucleic acid sequence of the gene is known, or the sequence of the nucleic acid complementary to the coding strand is available, or alternatively, if the target amino acid sequence is known, one may infer potential nucleic acid
15 sequences using known and preferred coding residues for each amino acid residue.

A preferred method of practicing this invention is to use carefully selected oligonucleotide sequences to screen cDNA libraries from various tissues, preferably human breast, colon, salivary gland, placental, fetal, brain, and carcinoma cell lines. Other biological sources of DNA encoding an heregulin-like ligand include other mammals and birds. Among the preferred mammals are members of the following
20 orders: bovine, ovine, equine, murine, and rodentia.

The oligonucleotide sequences selected as probes should be of sufficient length and sufficiently unambiguous that false positives are minimized. The actual nucleotide sequence(s) may, for example, be based on conserved or highly homologous nucleotide sequences or regions of HRG- α . The oligonucleotides may be degenerate at one or more positions. The use of degenerate oligonucleotides may be of particular
25 importance where a library is screened from a species in which preferential codon usage in that species is not known. The oligonucleotide must be labeled such that it can be detected upon hybridization to DNA in the library being screened. The preferred method of labeling is to use ^{32}P -labeled ATP with polynucleotide kinase, as is well known in the art, to radiolabel the oligonucleotide. However, other methods may be used to label the oligonucleotide, including, but not limited to, biotinylation or enzyme labeling.

30 Of particular interest is HRG- α nucleic acid that encodes a full-length polypeptide. In some preferred embodiments, the nucleic acid sequence includes the native HRG- α signal sequence. Nucleic acid having all the protein coding sequence is obtained by screening selected cDNA or genomic libraries, and, if necessary, using conventional primer extension procedures as described in section 7.79 of Sambrook *et al.*, *supra*, to detect precursors and processing intermediates of mRNA that may not have been reverse-
35 transcribed into cDNA.

HRG- α encoding DNA of Figures 1A-1D may be used to isolate DNA encoding the analogous ligand from other animal species via hybridization employing the methods discussed above. The preferred animals are mammals, particularly bovine, ovine, equine, feline, canine and rodentia, and more specifically rats, mice and rabbits.

40 **B. Amino Acid Sequence Variants of Heregulin**

Amino acid sequence variants of HRG are prepared by introducing appropriate nucleotide changes into HRG DNA, or by *in vitro* synthesis of the desired HRG polypeptide. Such variants include, for example, deletions from, or insertions or substitutions of, residues within the amino acid sequence shown for human HRG sequences. Any combination of deletion, insertion, and substitution can be made to arrive at the
45 final construct, provided that the final construct possesses the desired characteristics. Excluded from the

represent naturally occurring alleles (which will not require manipulation of HRG DNA) or predetermined mutant forms made by mutating the DNA, either to arrive at an allele or a variant not found in nature. In general, the location and nature of the mutation chosen will depend upon HRG characteristic to be modified. Obviously, such variations that, for example, convert HRG into a known receptor ligand, are not included within the scope of this invention, nor are any other HRG variants or polypeptide sequences that are not novel and unobvious over the prior art.

Amino acid sequence deletions generally range from about 1 to 30 residues, more preferably about 1 to 10 residues, and typically about 1 to 5 are contiguous. Deletions may be introduced into regions of low homology with other EGF family precursors to modify the activity of HRG. Deletions from HRG in areas of substantial homology with other EGF family sequences will be more likely to modify the biological activity of HRG more significantly. The number of consecutive deletions will be selected so as to preserve the tertiary structure of HRG in the affected domain, e.g., cysteine crosslinking, beta-pleated sheet or alpha helix.

Amino acid sequence insertions include amino- and/or carboxyl-terminal fusions ranging in length from one residue to polypeptides containing a hundred or more residues, as well as intrasequence insertions of single or multiple amino acid residues. Intrasequence insertions (i.e., insertions within HRG sequence) may range generally from about 1 to 10 residues, more preferably 1 to 5, and most preferably 1 to 3. Examples of terminal insertions include HRG with an N-terminal methionyl residue (an artifact of the direct expression of HRG in bacterial recombinant cell culture), and fusion of a heterologous N-terminal signal sequence to the N-terminus of HRG to facilitate the secretion of mature HRG from recombinant host cells. Such signal sequences generally will be obtained from, and thus be homologous to, the intended host cell species. Suitable sequences include STII, tPA or lpp for *E. coli*, alpha factor for yeast, and viral signals such as herpes gD for mammalian cells.

Other insertional variants of HRG include the fusion to the N- or C-terminus of HRG of an immunogenic polypeptide, e.g., bacterial polypeptides such as beta-lactamase or an enzyme encoded by the *E. coli trp* locus, or yeast protein, bovine serum albumin, and chemotactic polypeptides. C-terminal fusions of HRG-ECD with proteins having a long half-life such as immunoglobulin constant regions (or other immunoglobulin regions), albumin, or ferritin, as described in WO 89/02922, published 6 April 1989 are contemplated.

Another group of variants are amino acid substitution variants. These variants have at least one amino acid residue in HRG molecule removed and a different residue inserted in its place. The sites of greatest interest for substitutional mutagenesis include sites identified as the active site(s) of HRG, and sites where the amino acids found in HRG ligands from various species are substantially different in terms of side-chain bulk, charge, and/or hydrophobicity. A likely sub-domain of HRG-GFD having biological activity as a growth factor is the C-terminal segment, in particular within the sequence about from glycine 218 to valine 226 (HRG- α), and glycine 218 to lysine 228/serine 228 (HRG- β) based upon analogy to the EGF sub-sequence found to have EGF activity.

Other sites of interest are those in which particular residues of HRG-like ligands obtained from various species are identical. These positions may be important for the biological activity of HRG. These sites, especially those falling within a sequence of at least three other identically conserved sites, are substituted in a relatively conservative manner. Such conservative substitutions are shown in Table 1 under the heading of "preferred substitutions". If such substitutions result in a change in biological activity, then more substantial changes, denominated exemplary substitutions in Table 1, or as further described below in reference to amino acid classes, are introduced and the products screened.

In another embodiment, any methionyl residue other than the starting methionyl residue of the signal sequence, or any residue located within about three residues N- or C-terminal to each such methionyl residue, is substituted by another residue (preferably in accord with Table 1) or deleted. Alternatively, about 1-3 residues are inserted adjacent to such sites.

Any cysteine residues not involved in maintaining the proper conformation of HRG also may be substituted, generally with serine, to improve the oxidative stability of the molecule and prevent aberrant crosslinking.

Sites particularly suited for substitutions, deletions or insertions, or use as fragments, include, numbered from the N-terminus of HRG- α of Figure 1A - 1D:

- 1) potential glycosaminoglycan addition sites at the serine-glycine dipeptides at 42-43, 64-65, 151-152;
- 2) potential asparagine-linked glycosylation at positions 164, 170, 208 and 437, sites (NDS) 164-166, (NIT) 170-172, (NTS) 208-210, and NTS (609-611);
- 3) potential O-glycosylation in a cluster of serine and threonine at 209-218;
- 4) cysteines at 226, 234, 240, 254, 256 and 265;
- 5) transmembrane domain at 287-309;
- 6) loop 1 delineated by cysteines 226 and 240;
- 7) loop 2 delineated by cysteines 234 and 254;
- 8) loop 3 delineated by cysteines 256 and 265; and
- 9) potential protease processing sites at 2-3, 8-9, 23-24, 33-34, 36-37, 45-46, 48-49, 62-63, 66-67, 86-87, 110-111, 123-124, 134-135, 142-143, 272-273, 278-279 and 285-286;

Analogous regions in HRG- β 1 may be determined by reference to its' sequence. The analogous HRG- β 1 amino acids may be mutated or modified as discussed above for HRG- α . Analogous regions in HRG- β 2 may also be determined by reference to its' sequence. The analogous HRG- β 2 amino acids may be mutated or modified as discussed above for HRG- α or HRG- β 1. Analogous regions in HRG- β 3 may be determined by reference to its' sequence. Further, the analogous HRG- β 3 amino acids may be mutated or modified as discussed above for HRG- α , HRG- β 1, or HRG- β 2.

Another HRG variant is γ -HRG (or gamma-heregulin). γ -HRG is any polypeptide sequence that possesses at least one biological property of native sequence γ -HRG having SEQ ID NO:11. The biological property of this variant is the same as for HRG noted above. This variant encompasses not only the polypeptide isolated from a native γ -HRG source such as human MDA-MB-175 cells or from another source, such as another animal species, but also the polypeptide prepared by recombinant or synthetic methods. It also includes variant forms including functional derivatives, allelic variants, naturally occurring isoforms and analogues thereof. Sometimes the γ -HRG is "native γ -HRG" which refers to endogenous γ -HRG polypeptide which has been isolated from a mammal. The γ -HRG can also be "native sequence γ -HRG" insofar as it has the same amino acid sequence as a native γ -HRG (e.g. human γ -HRG shown in Fig. 7A-7C). Amino acid sequence variants of the native sequence are prepared by introducing appropriate nucleotide changes into the native sequence DNA, or by *in vitro* synthesis of the desired polypeptide. Such variants include, for example, deletions from, or insertions or substitutions of, residues within the amino acid sequence shown for the human protein in Fig. 7A-7C as generally described above for other HRG. Any combination of deletion, insertion, and substitution is made to arrive at the final construct, provided that the final construct possesses the desired characteristics. The amino acid changes also may alter post-translational processes of the native sequence, such as changing the number or position of O-linked

A183G, K185E, E186R, K187E, T188G, F197Y, M198R, D201T;
 A183G, K185E, E186R, K187E, T188G, P205Y, S206G, R207Y, Y208L, L209M;
 A183G, K185E, E186R, K187E, T188G, F197Y, M198R, D201T, P205Y, S206G, R207Y, Y208L, L209M;
 A183G, K185E, E186R, K187E, T188G, M226I;
 5 F197Y, M198R, D201T, P205Y, S206G, R207Y, Y208L, L209M;
 F197Y, M198R, D201T, P205Y, S206G, R207Y, Y208L, L209M, M226I;
 F197Y, M198R, D201T, M226I;
 A183G, K185E, E186R, K187E, T188G, F197Y, M198R, D201T, M226I;
 A183G, K185E, E186R, K187E, T188G, P205Y, S206G, R207Y, Y208L, L209M, M226I;
 10 A183G, K185E, E186R, K187E, T188G, F197Y, M198R, D201T, P205Y, S206G, R207Y, Y208L, L209M,
 M226I;
 F197Y, M198R, D201T, P205Y, S206G, R207Y, Y208L, L209M, N223H, M226I; and
 A183G, K185E, E186R, K187E, T188G, F197Y, M198R, D201T, P205Y, S206G, R207Y, Y208L, L209M,
 N223H, M226I.

15 In addition to including one or more of the amino acid substitutions disclosed herein, the heregulin
 variant can have one or more other modifications, such as an amino acid substitution, an insertion of at least
 one amino acid, a deletion of at least one amino acid, or a chemical modification. For example, the invention
 provides a heregulin variant that is a fragment. In a variation of this embodiment, the fragment includes
 residues corresponding to a portion of human heregulin- β 1 extending from about residue 175 to about
 20 residue 230 (i.e., the EGF-like domain). For example, the fragment can extend from residue 177 to residue
 244 and may be prepared by recombinant techniques (rHRG β 1-177-244).

DNA encoding amino acid sequence variants of HRG is prepared by a variety of methods known in
 the art. These methods include, but are not limited to, isolation from a natural source (in the case of naturally
 occurring amino acid sequence variants) or preparation by oligonucleotide-mediated (or site-directed)
 25 mutagenesis, PCR mutagenesis, and cassette mutagenesis of an earlier prepared variant or a non-variant
 version of HRG. These techniques may utilize HRG nucleic acid (DNA or RNA), or nucleic acid
 complementary to HRG nucleic acid.

Oligonucleotide-mediated mutagenesis is a preferred method for preparing substitution, deletion,
 and insertion variants of HRG DNA. This technique is well known in the art as described by Adelman *et al.*
 30 *DNA*, 2: 183 (1983). Briefly, HRG DNA is altered by hybridizing an oligonucleotide encoding the desired
 mutation to a DNA template, where the template is the single-stranded form of a plasmid or bacteriophage
 containing the unaltered or native DNA sequence of HRG. After hybridization, a DNA polymerase is used
 to synthesize an entire second complementary strand of the template that will thus incorporate the
 oligonucleotide primer, and will code for the selected alteration in HRG DNA.

35 Generally, oligonucleotides of at least 25 nucleotides in length are used. An optimal
 oligonucleotide will have 12 to 15 nucleotides that are completely complementary to the template on either
 side of the nucleotide(s) coding for the mutation. This ensures that the oligonucleotide will hybridize
 properly to the single-stranded DNA template molecule. The oligonucleotides are readily synthesized using
 techniques known in the art such as that described by Crea *et al.* (*Proc. Natl. Acad. Sci. USA*, 75:
 40 5765, 1978).

Single-stranded DNA template may also be generated by denaturing double-stranded plasmid (or
 other) DNA using standard techniques.

For alteration of the native DNA sequence (to generate amino acid sequence variants, for example),
 the oligonucleotide is hybridized to the single-stranded template under suitable hybridization conditions. A
 45 DNA polymerizing enzyme, usually the Klenow fragment of DNA polymerase I, is then added to synthesize
 the complementary strand of the template using the oligonucleotide as a primer for synthesis. A

primers is designed to overlap the position of the mutation and to contain the mutation: the sequence of the other primer must be identical to a stretch of sequence of the opposite strand of the plasmid, but this sequence can be located anywhere along the plasmid DNA. It is preferred, however, that the sequence of the second primer is located within 200 nucleotides from that of the first, such that in the end the entire amplified region of DNA bounded by the primers can be easily sequenced. PCR amplification using a primer pair like the one just described results in a population of DNA fragments that differ at the position of the mutation specified by the primer, and possibly at other positions, as template copying is somewhat error-prone.

If the ratio of template to product material is extremely low, the vast majority of product DNA fragments incorporate the desired mutation(s). This product material is used to replace the corresponding region in the plasmid that served as PCR template using standard DNA technology. Mutations at separate positions can be introduced simultaneously by either using a mutant second primer, or performing a second PCR with different mutant primers and ligating the two resulting PCR fragments simultaneously to the vector fragment in a three (or more)-part ligation.

In a specific example of PCR mutagenesis, template plasmid DNA (1mg) is linearized by digestion with a restriction endonuclease that has a unique recognition site in the plasmid DNA outside of the region to be amplified. Of this material, 100 ng is added to a PCR mixture containing PCR buffer, which contains the four deoxynucleotide tri-phosphates and is included in the GENEAMP kits (obtained from Perkin-Elmer Cetus, Norwalk, CT and Emeryville, CA), and 25 pmole of each oligonucleotide primer, to a final volume of 50 µl. The reaction mixture is overlaid with 35 µl mineral oil. The reaction is denatured for 5 minutes at 100°C, placed briefly on ice, and then 1 µl *Thermus aquaticus* (Taq) DNA polymerase (5 units/ml, purchased from Perkin-Elmer Cetus, Norwalk, CT and Emeryville, CA) is added below the mineral oil layer. The reaction mixture is then inserted into a DNA Thermal Cycler (purchased from Perkin-Elmer Cetus) programmed as follows:

2 min. 55°C,
30 sec. 72°C, then 19 cycles of the following:
30 sec. 94°C,
30 sec. 55°C, and
30 sec. 72°C.

At the end of the program, the reaction vial is removed from the thermal cycler and the aqueous phase transferred to a new vial, extracted with phenol/chloroform (50:50:vol), and ethanol precipitated, and the DNA is recovered by standard procedures. This material is subsequently subjected to the appropriate treatments for insertion into a vector.

Another method for preparing variants, cassette mutagenesis, is based on the technique described by Wells *et al.* (*Gene*, 34: 315,1985). The starting material is the plasmid (or other vector) comprising HRG DNA to be mutated. The codon(s) in HRG DNA to be mutated are identified. There must be a unique restriction endonuclease site on each side of the identified mutation site(s). If no such restriction sites exist, they may be generated using the above-described oligonucleotide-mediated mutagenesis method to introduce them at appropriate locations in HRG DNA. After the restriction sites have been introduced into the plasmid, the plasmid is cut at these sites to linearize it. A double-stranded oligonucleotide encoding the sequence of the DNA between the restriction sites but containing the desired mutation(s) is synthesized using standard procedures. The two strands are synthesized separately and then hybridized together using standard techniques. This double-stranded oligonucleotide is referred to as the cassette. This cassette is designed to have 3' and 5' ends that are compatible with the ends of the linearized plasmid, such that it can be directly ligated to the plasmid. This plasmid now contains the mutated HRG DNA sequence.

C. Insertion of DNA into a Cloning Vehicle

The cDNA or genomic DNA encoding native or variant HRG is inserted into a replicable vector for further cloning (amplification of the DNA) or for expression. Many vectors are available, and selection of

encoding HRG is more complex than that of an exogenously replicated vector because restriction enzyme digestion is required to excise HRG DNA. DNA can be amplified by PCR and directly transfected into the host cells without any replication component.

(iii) Selection Gene Component

5 Expression and cloning vectors should contain a selection gene, also termed a selectable marker. This gene encodes a protein necessary for the survival or growth of transformed host cells grown in a selective culture medium. Host cells not transformed with the vector containing the selection gene will not survive in the culture medium. Typical selection genes encode proteins that (a) confer resistance to antibiotics or other toxins, e.g., ampicillin, neomycin, methotrexate, or tetracycline, (b) complement
10 auxotrophic deficiencies, or (c) supply critical nutrients not available from complex media, e.g., the gene encoding D-alanine racemase for *Bacilli*.

One example of a selection scheme utilizes a drug to arrest growth of a host cell. Those cells that are successfully transformed with a heterologous gene express a protein conferring drug resistance and thus survive the selection regimen. Examples of such dominant selection use the drugs neomycin (Southern *et al.*, *J. Molec. Appl. Genet.* 1: 327,1982), mycophenolic acid (Mulligan *et al.*, *Science* 209: 1422,1980) or
15 hygromycin (Sugden *et al.*, *Mol. Cell. Biol.* 5: 410-413,1985). The three examples given above employ bacterial genes under eukaryotic control to convey resistance to the appropriate drug G418 or neomycin (geneticin), xgpt (mycophenolic acid), or hygromycin, respectively.

Another example of suitable selectable markers for mammalian cells are those that enable the
20 identification of cells competent to take up HRG nucleic acid, such as dihydrofolate reductase (DHFR) or thymidine kinase. The mammalian cell transformants are placed under selection pressure which only the transformants are uniquely adapted to survive by virtue of having taken up the marker. Selection pressure is imposed by culturing the transformants under conditions in which the concentration of selection agent in the medium is successively changed, thereby leading to amplification of both the selection gene and the DNA
25 that encodes HRG. Amplification is the process by which genes in greater demand for the production of a protein critical for growth are reiterated in tandem within the chromosomes of successive generations of recombinant cells. Increased quantities of HRG are synthesized from the amplified DNA.

For example, cells transformed with the DHFR selection gene are first identified by culturing all of the transformants in a culture medium that contains methotrexate (Mtx), a competitive antagonist of DHFR.
30 An appropriate host cell when wild-type DHFR is employed is the Chinese hamster ovary (CHO) cell line deficient in DHFR activity, prepared and propagated as described by Urlaub and Chasin, *Proc. Natl. Acad. Sci. USA*, 77: 4216, 1980. The transformed cells are then exposed to increased levels of methotrexate. This leads to the synthesis of multiple copies of the DHFR gene, and, concomitantly, multiple copies of other DNA comprising the expression vectors, such as the DNA encoding HRG. This amplification technique can
35 be used with any otherwise suitable host, e.g., ATCC No. CCL61 CHO-K1, notwithstanding the presence of endogenous DHFR if, for example, a mutant DHFR gene that is highly resistant to Mtx is employed (EP 117,060). Alternatively, host cells (particularly wild-type hosts that contain endogenous DHFR) transformed or co-transformed with DNA sequences encoding HRG, wild-type DHFR protein, and another selectable marker such as aminoglycoside 3' phosphotransferase (APH) can be selected by cell growth in medium
40 containing a selection agent for the selectable marker such as an aminoglycosidic antibiotic, e.g., kanamycin, neomycin, or G418 (see U.S. Pat. No. 4,965,199).

A suitable selection gene for use in yeast is the *trp1* gene present in the yeast plasmid YRp7 (Stinchcomb *et al.*, *Nature*, 282: 39, 1979; Kingsman *et al.*, *Gene*, 7: 141, 1979; or Tschemper *et al.*, *Gene*,
45 10: 157, 1980). The *trp1* gene provides a selection marker for a mutant strain of yeast lacking the ability to grow in tryptophan; for example, ATCC No. 44076 or PEP4-1 (Jones, *Genetics*, 85: 12, 1977). The presence of the *trp1* lesion in the yeast host cell genome then provides an effective environment for detecting

promoters. e.g., the actin promoter or an immunoglobulin promoter, from heat-shock promoters, and from the promoter normally associated with HRG sequence, provided such promoters are compatible with the host cell systems.

The early and late promoters of the SV40 virus are conveniently obtained as an SV40 restriction fragment that also contains the SV40 viral origin of replication (Fiers *et al.*, *Nature*, 273:113 (1978); Mulligan and Berg, *Science*, 209: 1422-1427 (1980); Pavlakis *et al.*, *Proc. Natl. Acad. Sci. USA*, 78: 7398-7402 (1981)). The immediate early promoter of the human cytomegalovirus is conveniently obtained as a *HindIII* E restriction fragment (Greenaway *et al.*, *Gene*, 18: 355-360 (1982)). A system for expressing DNA in mammalian hosts using the bovine papilloma virus as a vector is disclosed in U.S. 4,419,446. A modification of this system is described in U.S. 4,601,978. See also Gray *et al.*, *Nature*, 295: 503-508 (1982) on expressing cDNA encoding immune interferon in monkey cells; Reyes *et al.*, *Nature*, 297: 598-601 (1982) on expression of human b-interferon cDNA in mouse cells under the control of a thymidine kinase promoter from herpes simplex virus; Canaani and Berg, *Proc. Natl. Acad. Sci. USA*, 79: 5166-5170 (1982) on expression of the human interferon b1 gene in cultured mouse and rabbit cells; and Gorman *et al.*, *Proc. Natl. Acad. Sci. USA*, 79: 6777-6781 (1982) on expression of bacterial CAT sequences in CV-1 monkey kidney cells, chicken embryo fibroblasts, Chinese hamster ovary cells, HeLa cells, and mouse NIH-3T3 cells using the Rous sarcoma virus long terminal repeat as a promoter.

(v) Enhancer Element Component

Transcription of a DNA encoding HRG of this invention by higher eukaryotes is often increased by inserting an enhancer sequence into the vector. Enhancers are cis-acting elements of DNA, usually about from 10-300 bp, that act on a promoter to increase its transcription. Enhancers are relatively orientation and position independent having been found 5' (Laimins *et al.*, *Proc. Natl. Acad. Sci. USA*, 78: 993, 1981) and 3' (Lusky *et al.*, *Mol. Cell Bio.*, 3: 1108, 1983) to the transcription unit, within an intron (Banerji *et al.*, *Cell*, 33: 729, 1983) as well as within the coding sequence itself (Osborne *et al.*, *Mol. Cell Bio.*, 4: 1293, 1984). Many enhancer sequences are now known from mammalian genes (globin, elastase, albumin, a-fetoprotein and insulin). Typically, however, one will use an enhancer from a eukaryotic cell virus. Examples include the SV40 enhancer on the late side of the replication origin (bp 100-270), the cytomegalovirus early promoter enhancer, the polyoma enhancer on the late side of the replication origin, and adenovirus enhancers (see also Yaniv, *Nature*, 297: 17-18 (1982)) on enhancing elements for activation of eukaryotic promoters. The enhancer may be spliced into the vector at a position 5' or 3' to HRG DNA, but is preferably located at a site 5' from the promoter.

(vi) Transcription Termination Component

Expression vectors used in eukaryotic host cells (yeast, fungi, insect, plant, animal, human, or nucleated cells from other multicellular organisms) will also contain sequences necessary for the termination of transcription and for stabilizing the mRNA. Such sequences are commonly available from the 5' and, occasionally 3' untranslated regions of eukaryotic or viral DNAs or cDNAs. These regions contain nucleotide segments transcribed as polyadenylated fragments in the untranslated portion of the mRNA encoding HRG. The 3' untranslated regions also include transcription termination sites.

Construction of suitable vectors containing one or more of the above listed components the desired coding and control sequences employs standard ligation techniques. Isolated plasmids or DNA fragments are cleaved, tailored, and religated in the form desired to generate the plasmids required.

For analysis to confirm correct sequences in plasmids constructed, the ligation mixtures are used to transform *E. coli* K12 strain 294 (ATCC 31,446) and successful transformants selected by ampicillin or tetracycline resistance where appropriate. Plasmids from the transformants are prepared, analyzed by restriction endonuclease digestion, and/or sequenced by the method of Messing *et al.*, *Nucleic Acids Res.* 9: 309 (1981) or by the method of Maxam *et al.*, *Methods in Enzymology* 65: 499 (1980).

Plant cell cultures of cotton, corn, potato, soybean, petunia, tomato, and tobacco can be utilized as hosts. Typically, plant cells are transfected by incubation with certain strains of the bacterium *Agrobacterium tumefaciens*, which has been previously manipulated to contain HRG DNA. During incubation of the plant cell culture with *A. tumefaciens*, the DNA encoding HRG is transferred to the plant cell host such that it is transfected, and will, under appropriate conditions, express HRG DNA. In addition, regulatory and signal sequences compatible with plant cells are available, such as the nopaline synthase promoter and polyadenylation signal sequences (Depicker *et al.*, *J. Mol. Appl. Gen.*, 1: 561 (1982)). In addition, DNA segments isolated from the upstream region of the T-DNA 780 gene are capable of activating or increasing transcription levels of plant-expressible genes in recombinant DNA-containing plant tissue (see EP 321,196, published 21 June 1989).

However, interest has been greatest in vertebrate cells, and propagation of vertebrate cells in culture (tissue culture) has become a routine procedure in recent years (*Tissue Culture*, Academic Press, Kruse and Patterson, editors (1973)). Examples of useful mammalian host cell lines are monkey kidney CV1 line transformed by SV40 (COS-7, ATCC CRL 1651); human embryonic kidney line (293 or 293 cells subcloned for growth in suspension culture, Graham *et al.*, *J. Gen. Virol.*, 36: 59, 1977); baby hamster kidney cells (BHK, ATCC CCL 10); Chinese hamster ovary cells/-DHFR (CHO, Urlaub and Chasin, *Proc. Natl. Acad. Sci. USA*, 77: 4216 (1980)); mouse sertoli cells (TM4, Mather, *Biol. Reprod.*, 23: 243-251 (1980)); monkey kidney cells (CV1 ATCC CCL 70); African green monkey kidney cells (VERO-76, ATCC CRL-1587); human cervical carcinoma cells (HELA, ATCC CCL 2); canine kidney cells (MDCK, ATCC CCL 34); buffalo rat liver cells (BRL 3A, ATCC CRL 1442); human lung cells (W138, ATCC CCL 75); human liver cells (Hep G2, HB 8065); mouse mammary tumor (MMT 060562, ATCC CCL51); TRI cells (Mather *et al.*, *Annals N.Y. Acad. Sci.*, 383: 44-68 (1982)); MRC 5 cells; FS4 cells; and a human hepatoma cell line (Hep G2). Preferred host cells are human embryonic kidney 293 and Chinese hamster ovary cells.

Host cells are transfected and preferably transformed with the above-described expression or cloning vectors of this invention and cultured in conventional nutrient media modified as appropriate for inducing promoters, selecting transformants, or amplifying the genes encoding the desired sequences.

Transfection refers to the taking up of an expression vector by a host cell whether or not any coding sequences are in fact expressed. Numerous methods of transfection are known to the ordinarily skilled artisan, for example, CaPO_4 and electroporation. Successful transfection is generally recognized when any indication of the operation of this vector occurs within the host cell.

Transformation means introducing DNA into an organism so that the DNA is replicable, either as an extrachromosomal element or by chromosomal integration. Depending on the host cell used, transformation is done using standard techniques appropriate to such cells. The calcium treatment employing calcium chloride, as described in section 1.82 of Sambrook *et al.*, *supra*, is generally used for prokaryotes or other cells that contain substantial cell-wall barriers. Infection with *Agrobacterium tumefaciens* is used for transformation of certain plant cells, as described by Shaw *et al.*, *Gene*, 23: 315 (1983) and WO 89/05859, published 29 June 1989. For mammalian cells without such cell walls, the calcium phosphate precipitation method described in sections 16.30-16.37 of Sambrook *et al.*, *supra*, is preferred. General aspects of mammalian cell host system transformations have been described by Axel in U.S. 4,399,216, issued 16 August 1983. Transformations into yeast are typically carried out according to the method of Van Solingen *et al.*, *J. Bact.*, 130: 946 (1977) and Hsiao *et al.*, *Proc. Natl. Acad. Sci. (USA)*, 76: 3829 (1979). However, other methods for introducing DNA into cells such as by nuclear injection, electroporation, or protoplast fusion may also be used.

E. Culturing the Host Cells

Prokaryotic cells used to produce HRG polypeptide of this invention are cultured in suitable media as described generally in Sambrook *et al.*, *supra*.

prepared against a native HRG polypeptide or against a synthetic peptide based on the DNA sequences provided herein as described further below.

G. Purification of The Heregulin Polypeptides

HRG is recovered from a cellular membrane fraction. Alternatively, a proteolytically cleaved or a truncated expressed soluble HRG fragment or subdomain are recovered from the culture medium as a soluble polypeptide. A HRG is recovered from host cell lysates when directly expressed without a secretory signal.

When HRG is expressed in a recombinant cell other than one of human origin, HRG is completely free of proteins or polypeptides of human origin. However, it is desirable to purify HRG from recombinant cell proteins or polypeptides to obtain preparations that are substantially homogeneous as to HRG. As a first step, the culture medium or lysate is centrifuged to remove particulate cell debris. The membrane and soluble protein fractions are then separated. HRG is then be purified from both the soluble protein fraction (requiring the presence of a protease) and from the membrane fraction of the culture lysate, depending on whether HRG is membrane bound. The following procedures are exemplary of suitable purification procedures: fractionation on immunoaffinity or ion-exchange columns; ethanol precipitation; reverse phase HPLC; chromatography on silica, heparin SEPHAROSE or on a cation exchange resin such as DEAE; chromatofocusing; SDS-PAGE; ammonium sulfate precipitation; and gel filtration using, for example, SEPHADEX G-75.

HRG variants in which residues have been deleted, inserted or substituted are recovered in the same fashion as the native HRG, taking account of any substantial changes in properties occasioned by the variation. For example, preparation of a HRG fusion with another protein or polypeptide, e.g., a bacterial or viral antigen, facilitates purification; an immunoaffinity column containing antibody to the antigen can be used to adsorb the fusion. Immunoaffinity columns such as a rabbit polyclonal anti-HRG column can be employed to absorb HRG variant by binding it to at least one remaining immune epitope. A protease inhibitor such as phenylmethylsulfonylfluoride (PMSF) also may be useful to inhibit proteolytic degradation during purification, and antibiotics may be included to prevent the growth of adventitious contaminants. One skilled in the art will appreciate that purification methods suitable for native HRG may require modification to account for changes in the character of HRG variants or upon expression in recombinant cell culture.

H. Covalent Modifications of HRG

Covalent modifications of HRG polypeptides are included within the scope of this invention. Both native HRG and amino acid sequence variants of HRG optionally are covalently modified. One type of covalent modification included within the scope of this invention is a HRG polypeptide fragment. HRG fragments, such as HRG-GDF, having up to about 40 amino acid residues are conveniently prepared by chemical synthesis, or by enzymatic or chemical cleavage of the full-length HRG polypeptide or HRG variant polypeptide. Other types of covalent modifications of HRG or fragments thereof are introduced into the molecule by reacting targeted amino acid residues of HRG or fragments thereof with an organic derivatizing agent that is capable of reacting with selected side chains or the N- or C-terminal residues.

Cysteiny residues most commonly are reacted with α -haloacetates (and corresponding amines), such as chloroacetic acid or chloroacetamide, to give carboxymethyl or carboxyamidomethyl derivatives. Cysteiny residues also are derivatized by reaction with bromotrifluoroacetone, α -bromo- β -(5-imidozoyl)propionic acid, chloroacetyl phosphate, N-alkylmaleimides, 3-nitro-2-pyridyl disulfide, methyl 2-pyridyl disulfide, p-chloromercuribenzoate, 2-chloromercuri-4-nitrophenol, or chloro-7-nitrobenzo-2-oxa-1,3-diazole.

Histidyl residues are derivatized by reaction with diethylpyrocarbonate at pH 5.5-7.0 because this agent is relatively specific for the histidyl side chain. Para-bromophenacyl bromide also is useful; the reaction is preferably performed in 0.1M sodium cacodylate at pH 6.0.

asparagine-X-serine and asparagine-X-threonine, where X is any amino acid except proline, are the recognition sequences for enzymatic attachment of the carbohydrate moiety to the asparagine side chain. Thus, the presence of either of these tri-peptide sequences in a polypeptide creates a potential glycosylation site. O-linked glycosylation refers to the attachment of one of the sugars N-acetylgalactosamine, galactose, or xylose, to a hydroxyamino acid, most commonly serine or threonine, although 5-hydroxyproline or 5-hydroxylysine may also be used.

Glycosylation sites are added to HRG by altering its amino acid sequence to contain one or more of the above-described tri-peptide sequences (for N-linked glycosylation sites). The alteration may also be made by the addition of, or substitution by, one or more serine or threonine residues to HRG (for O-linked glycosylation sites). For ease, HRG is preferably altered through changes at the DNA level, particularly by mutating the DNA encoding HRG at preselected bases such that codons are generated that will translate into the desired amino acids.

Chemical or enzymatic coupling of glycosides to HRG increases the number of carbohydrate substituents. These procedures are advantageous in that they do not require production of the polypeptide in a host cell that is capable of N- and O-linked glycosylation. Depending on the coupling mode used, the sugar(s) may be attached to (a) arginine and histidine, (b) free carboxyl groups, (c) free sulfhydryl groups such as those of cysteine, (d) free hydroxyl groups such as those of serine, threonine, or hydroxyproline, (e) aromatic residues such as those of phenylalanine, tyrosine, or tryptophan, or (f) the amide group of glutamine. These methods are described in WO 87/05330, published 11 September 1987, and in Aplin and Wriston (*CRC Crit. Rev. Biochem.*, pp. 259-306 (1981)).

Carbohydrate moieties present on an HRG also are removed chemically or enzymatically. Chemical deglycosylation requires exposure of the polypeptide to the compound trifluoromethanesulfonic acid, or an equivalent compound. This treatment results in the cleavage of most or all sugars except the linking sugar (N-acetylglucosamine or N-acetylgalactosamine), while leaving the polypeptide intact. Chemical deglycosylation is described by Hakimuddin *et al.* (*Arch. Biochem. Biophys.*, 259:52 (1987)) and by Edge *et al.* (*Anal. Biochem.*, 118:131 (1981)). Carbohydrate moieties are removed from HRG by a variety of endo- and exo- glycosidases as described by Thotakura *et al.* (*Meth. Enzymol.*, 138:350 (1987)).

Glycosylation also is suppressed by tunicamycin as described by Duskin *et al.* (*J. Biol. Chem.*, 257:3105 (1982)). Tunicamycin blocks the formation of protein-N-glycoside linkages.

HRG may also be modified by linking HRG to various nonproteinaceous polymers, e.g., polyethylene glycol, polypropylene glycol or polyoxyalkylenes, in the manner set forth in U.S. 4,640,835; 4,496,689; 4,301,144; 4,670,417; 4,791,192 or 4,179,337.

One preferred way to increase the *in vivo* circulating half life of non-membrane bound HRG is to conjugate it to a polymer that confers extended half-life, such as polyethylene glycol (PEG). (Maxfield, *et al.*, *Polymer* 16,505-509 (1975); Bailey, F. E., *et al.*, in *Nonionic Surfactants* (Schick, M. J., ed.) pp.794-821, 1967); (Abuchowski, A. *et al.*, *J. Biol. Chem.* 252, 3582-3586, 1977; Abuchowski, A. *et al.*, *Cancer Biochem. Biophys.* 7, 175-186, 1984); (Katre, N.V. *et al.*, *Proc. Natl. Acad. Sci.*, 84, 1487-1491, 1987; Goodson, R. *et al. Bio Technology*, 8, 343-346, 1990). Conjugation to PEG also has been reported to have reduced immunogenicity and toxicity (Abuchowski, A. *et al.*, *J. Biol. Chem.*, 252, 3578-3581, 1977).

HRG may also be entrapped in microcapsules prepared, for example, by coacervation techniques or by interfacial polymerization (for example, hydroxymethylcellulose or gelatin-microcapsules and poly-(methylmethacrylate) microcapsules, respectively), in colloidal drug delivery systems (for example, liposomes, albumin microspheres, microemulsions, nano-particles and nanocapsules), or in macroemulsions. Such techniques are disclosed in *Remington's Pharmaceutical Sciences*, 16th edition, Osol, A., Ed., (1980).

Those skilled in the art will be capable of screening variants in order to select the optimal variant for the purpose intended. For example, a change in the immunological character of HRG, such as a change in affinity for a given antigen or for the HER2 receptor, is measured by a competitive-type immunoassay using

Culture medium in which hybridoma cells are growing is assayed for production of monoclonal antibodies directed against the antigen. Preferably, the binding specificity of monoclonal antibodies produced by hybridoma cells is determined by immunoprecipitation or by an *in vitro* binding assay, such as radioimmunoassay (RIA) or enzyme-linked immunoabsorbent assay (ELISA).

5 The binding affinity of the monoclonal antibody can, for example, be determined by the Scatchard analysis of Munson *et al.*, *Anal. Biochem.*, 107:220 (1980).

After hybridoma cells are identified that produce antibodies of the desired specificity, affinity, and/or activity, the clones may be subcloned by limiting dilution procedures and grown by standard methods (Goding, *Monoclonal Antibodies: Principles and Practice*, pp.59-103 (Academic Press, 1986)). Suitable
10 culture media for this purpose include, for example, D-MEM or RPMI-1640 medium. In addition, the hybridoma cells may be grown *in vivo* as ascites tumors in an animal.

The monoclonal antibodies secreted by the subclones are suitably separated from the culture medium, ascites fluid, or serum by conventional immunoglobulin purification procedures such as, for example, protein A-SEPHAROSE, hydroxylapatite chromatography, gel electrophoresis, dialysis, or affinity chromatography.

15 DNA encoding the monoclonal antibodies is readily isolated and sequenced using conventional procedures (e.g., by using oligonucleotide probes that are capable of binding specifically to genes encoding the heavy and light chains of the monoclonal antibodies). The hybridoma cells serve as a preferred source of such DNA. Once isolated, the DNA may be placed into expression vectors, which are then transfected into host cells such as *E. coli* cells, simian COS cells, Chinese hamster ovary (CHO) cells, or myeloma cells that
20 do not otherwise produce immunoglobulin protein, to obtain the synthesis of monoclonal antibodies in the recombinant host cells. Recombinant production of antibodies will be described in more detail below.

Hybridoma cell lines producing antibodies are identified by screening the culture supernatants for antibody which binds to HER2/HER3 receptors. This is routinely accomplished by conventional immunoassays using soluble receptor preparations or by FACS using cell-bound receptor and labeled
25 candidate antibody. Agonist antibodies are preferably antibodies which stimulate autophosphorylation in the HRG tyrosine autophosphorylation assay described above.

The hybrid cell lines can be maintained in culture *in vitro* in cell culture media. The cell lines of this invention can be selected and/or maintained in a composition comprising the continuous cell line in hypoxanthine-aminopterin thymidine (HAT) medium. In fact, once the hybridoma cell line is established, it
30 can be maintained on a variety of nutritionally adequate media. Moreover, the hybrid cell lines can be stored and preserved in any number of conventional ways, including freezing and storage under liquid nitrogen. Frozen cell lines can be revived and cultured indefinitely with resumed synthesis and secretion of monoclonal antibody. The secreted antibody is recovered from tissue culture supernatant by conventional methods such as precipitation, ion exchange chromatography, affinity chromatography, or the like. The
35 antibodies described herein are also recovered from hybridoma cell cultures by conventional methods for purification of IgG or IgM as the case may be that heretofore have been used to purify these immunoglobulins from pooled plasma, e.g., ethanol or polyethylene glycol precipitation procedures.

Human antibodies may be used and are preferable. Such antibodies can be obtained by using human hybridomas (Cote *et al.*, *Monoclonal Antibodies and Cancer Therapy*, Alan R. Liss, p. 77 (1985)).
40 Chimeric antibodies, Cabilly *et al.*, U.S. 4,816,567, (Morrison *et al.*, *Proc. Natl. Acad. Sci.*, 81:6851 (1984); Neuberger *et al.*, *Nature* 312:604 (1984); Takeda *et al.*, *Nature* 314:452 (1985)) containing a murine anti-HER2/HER3 variable region and a human constant region of appropriate biological activity (such as ability to activate human complement and mediate ADCC) are within the scope of this invention, as are humanized antibodies produced by conventional CDR-grafting methods (Riechmann *et al.*, *Nature* 332:333-327(1988);
45 EP 0328404 A1; EP 02394000 A2).

Techniques for creating recombinant DNA versions of the antigen-binding regions of antibody molecules (Fab or variable regions fragments) which bypass the generation of monoclonal antibodies are also

Therapeutic formulations of HRG or agonist antibody are prepared for storage by mixing the HRG protein having the desired degree of purity with optional physiologically acceptable carriers, excipients, or stabilizers (*Remington's Pharmaceutical Sciences, supra*), in the form of lyophilized cake or aqueous solutions. Acceptable carriers, excipients or stabilizers are nontoxic to recipients at the dosages and concentrations employed, and include buffers such as phosphate, citrate, and other organic acids; antioxidants including ascorbic acid; low molecular weight (less than about 10 residues) polypeptides; proteins, such as serum albumin, gelatin, or immunoglobulins; hydrophilic polymers such as polyvinylpyrrolidone; amino acids such as glycine, glutamine, asparagine, arginine or lysine; monosaccharides, disaccharides, and other carbohydrates including glucose, mannose, or dextrans; chelating agents such as EDTA; sugar alcohols such as mannitol or sorbitol; salt-forming counterions such as sodium; and/or nonionic surfactants such as TWEEN, PLURONICS or polyethylene glycol (PEG).

HRG or agonist antibody to be used for *in vivo* administration must be sterile. This is readily accomplished by filtration through sterile filtration membranes, prior to or following lyophilization and reconstitution. The HRG or antibody ordinarily will be stored in lyophilized form or in solution.

Therapeutic HRG or antibody compositions generally are placed into a container having a sterile access port, for example, an intravenous solution bag or vial having a stopper pierceable by a hypodermic injection needle.

The route of HRG or antibody administration is in accord with known methods, e.g., injection or infusion by intravenous, intraperitoneal, intracerebral, intramuscular, intraocular, intraarterial, powder or liquid aerosol administration to the nose or lung or intralesional routes, or by sustained release systems as noted below. The HRG ligand may be administered continuously by infusion or by bolus injection. An agonist antibody is preferably administered in the same fashion, or by administration into the blood stream or lymph.

The HRG, HRG variant or fragment and agonist antibodies may be spray dried or spray freeze dried using known techniques (Yeo et al, *Biotech. and Bioeng.*, 41:341-346 (1993); Gombotz et al, PCT/US90/02421).

Suitable examples of sustained-release preparations include semipermeable matrices of solid hydrophobic polymers containing the protein, which matrices are in the form of shaped articles, e.g. films, or microcapsules. Examples of sustained-release matrices include polyesters, hydrogels (e.g., poly(2-hydroxyethyl-methacrylate) as described by Langer et al., *J. Biomed. Mater. Res.*, 15: 167-277 (1981) and Langer, *Chem. Tech.*, 12: 98-105 (1982) or poly(vinylalcohol)), polylactides (U.S. 3,773,919, EP 58,481), copolymers of L-glutamic acid and gamma ethyl-L-glutamate (Sidman et al., *Biopolymers*, 22: 547-556 (1983)), non-degradable ethylene-vinyl acetate (Langer et al., *supra*), degradable lactic acid-glycolic acid copolymers such as the LUPRON DEPOT (injectable microspheres composed of lactic acid-glycolic acid copolymer and leuprolide acetate), and poly-D-(-)-3-hydroxybutyric acid (EP 133,988). While polymers such as ethylene-vinyl acetate and lactic acid-glycolic acid enable release of molecules for over 100 days, certain hydrogels release proteins for shorter time periods. When encapsulated proteins remain in the body for a long time, they may denature or aggregate as a result of exposure to moisture at 37°C, resulting in a loss of biological activity and possible changes in immunogenicity. Rational strategies can be devised for protein stabilization depending on the mechanism involved. For example, if the aggregation mechanism is discovered to be intermolecular S-S bond formation through thio-disulfide interchange, stabilization may be achieved by modifying sulfhydryl residues, lyophilizing from acidic solutions, controlling moisture content, using appropriate additives, and developing specific polymer matrix compositions.

Sustained-release HRG or antibody compositions also include liposomally entrapped HRG or antibody. Liposomes containing HRG or antibody are prepared by methods known *per se*: DE 3,218,121; Epstein et al., *Proc. Natl. Acad. Sci. USA*, 82: 3688-3692 (1985); Hwang et al., *Proc. Natl. Acad. Sci. USA*,

L⁶³⁶HER3-(TR)-DKTH²²⁴_{VH}; for HER4, G⁶⁴⁰HER4-(TR)-DKTH²²⁴_{VH}. The conserved TR sequence is derived from the MI I site. The final expression constructs were in a pRK-type plasmid backbone wherein eukaryotic expression is driven by a CMV promoter (Gorman *et al.*, *DNA Prot. Eng. Tech.* 2:3-10 (1990)).

To obtain protein for *in vitro* experiments, adherent HEK-293 cells were transfected with the appropriate expression plasmids using standard calcium phosphate methods (Gorman *et al.*, *supra* and Huang *et al.*, *Nucleic Acids Res.* 18:937-947 (1990)). Serum-containing media was replaced with serum-free media 15 hours post-transfection and the transfected cells incubated for 5-7 days. The resulting conditioned media was harvested and passed through Protein A columns (1 mL Pharmacia HiTrap™). Purified IgG fusions were eluted with 0.1 M citric acid (pH 4.2) into tubes containing 1 M Tris pH 9.0. The eluted proteins were subsequently dialyzed against PBS and concentrated using Centri-prep-30 filters (Amicon). Glycerol was added to a final concentration of 25% and the material stored at -20 C. Concentrations of material were determined via a Fc-ELISA.

Cell Culture: Human breast cancer cell lines MDA-MB-175, MDA-MB-231, SK-BR-3 and MCF7 were obtained from the American Type Culture Collection and maintained in a 50:50 mixture of F12 Ham's and Dulbecco's modified Eagle medium (DMEM), supplemented with 10% heat inactivated FBS, 2 mM glutamine and 10% penicillin-streptomycin.

Generation and Characterization of cDNA Library: Total RNA was purified from MDA-MB-175 cells using the guanidinium isothiocyanate-cesium chloride procedure (Sambrook *et al.*, *Molecular Cloning: A Laboratory Manual* (New York: Cold Spring Harbor Laboratory Press, (1989)). Poly (A)⁺ RNA was isolated using oligo (dT) Dynabeads (DYNAL) as recommended by the supplier. First and second strand syntheses were performed using a Gibco BRL cDNA synthesis kit. λ gt10 cDNA recombinants were generated when a cDNA cloning system from Amersham was used. *In vitro* packaging was performed using Gigapack II packaging extract (Stratagene). PstI-XhoI HRG β 3 cDNA fragment (nt 144-618) was labeled by random priming and 1 x 10⁶ plaques were screened. Positive clones were confirmed and purified by secondary and tertiary screening. Phage DNA was isolated as a BamHI fragment and subcloned into the corresponding site of pBluescript SK⁺. Clone 5 was completely sequenced using the Sequenase version 2.0 DNA sequencing kit (United States Biochemicals, Inc.). Both strands were sequenced.

Bacterial Expression System: A cDNA fragment of clone 5 (nt 1690- 2722) was subcloned into the pET-32 TRX fusion vector (Novagen). This BglII-BglII fragment was inserted into the BamHI site of the pET32a plasmid. The trxy-HRG (amino acids 455-768) protein expression in *E.coli* was induced as recommended by the supplier.

Purification of Recombinant γ -HRG: *E.coli* cells expressing trxy-HRG were collected and suspended at 9 ml/g in 50 mM Tris HCL pH 8. Lysozyme was added to a final concentration of 0.2 mg/ml and the solution was stirred on ice for 1 hr. Dnase I (10 μ g/ml) and MgCl₂ (4 mM) were added. The solution was then sonicated for 30 min and cell pellets collected afterwards. The pellet fraction was dissolved at 250 ml/g in 6 M Gdn HCL, 0.1 M Tris HCL, pH 8.8. Solubilized proteins were sulfitolyzed by adding 1/10 volume of 1 M Na₂SO₃ and 1/10 volume of 0.2 M Na₂S₄O₆. The reaction was allowed to proceed for 1.5 hours at room temperature and protein was purified by gel filtration chromatography using a High Load Superdex™ 75 prep grade column (Pharmacia). Refolding was initiated by the addition of 1 mM cysteine, and 10 mM methionine was added as an antioxidant and incubated overnight at room temperature. Protein concentration was determined by quantitative amino acid analysis.

Northern and Southern Hybridization: Total RNA was isolated by the method of Chomczynski *et al.* *Anal Biochem.* 162:156-159 (1987). Poly (A)⁺ was isolated using oligo d(T) cellulose columns (Qiagen) as recommended by the supplier. RNA was denatured and size fractioned in a 0.8% formaldehyde/ 1%

seeded in T175 flasks and grown until reaching 70-80% confluency ($\sim 2.5 \times 10^7$ cells/ flask). Subsequently, cells were washed with PBS and grown in serum free F12/ DMEM medium for 3-4 days. Medium was then collected, filtered and concentrated using an ultrafiltration cell with YM10 Diaflo ultrafiltration membranes (Amicon). γ -HRG was visualized in conditioned medium of MDA-MB-175 cells by Western blot analysis under non reducing conditions. γ -HRG was partially purified by HPLC using a C4 reverse phase column. CHO expressed full length HRG β 1 (lane 1) and semi pure γ -HRG (lane 2) were electrophoresed, blot was probed with HER2/HER4 IgG heterodimers and Western blot was developed. A ~ 64 kDa band could be seen in the lane containing partial purified supernatant whereas CHO expressed full length HRG β 1 migrated as a 45 kDa protein.

Cell Proliferation Assay with Crystal Violet: Tumor cell lines were plated in 96 well plates at following densities: 2×10^4 cells/well for MDA-MB-175 and 1×10^4 cells/well for SK-BR-3. The media contained 1% FBS and cells were allowed to adhere for 2 hours. Monoclonal antibodies, immunoadhesions (10 μ g/ml) or media alone were added and the cells were incubated for 2 hours at 37°C. rHRG β 1₁₇₇₋₂₄₄ was added at a final concentration of 1 nM, or 100 nM for neutralising the immunoadhesion, and the cells were incubated for 4 days. Monolayers were washed with PBS and stained/fixed with 0.5% crystal violet. Plates were air dried, the dye was eluted with 0.1 M sodium citrate (pH 4.2) in ethanol (50:50) and the absorbance was measured at 540 nm.

Isolation and sequence analysis of γ -HRG: To characterize the heregulin transcript in MDA-MB-175 cells, a λ gt 10 cDNA library was constructed with mRNA derived from this cell line. The library was screened with a cDNA probe corresponding to the EGF-like domain and part of the N-terminal sequence of HRG β 3. Various clones were identified. One of the clones which appeared to contain the full length cDNA sequence was isolated and sequenced. Fig.7A-7D shows the nucleotide sequence and the predicted amino acid sequence of γ -HRG. The single open reading frame of 2303 bp starts with an ATG codon at nt 334. This start codon lies in a nucleotide sequence context, which is known to be a potential translation initiation site (Kozak, *Nucleic Acid Research* 15:8125-8148 (1987)). Several termination codons were found upstream of the initiation codon. The stop codon TAG at nt 2637 is followed by the 3' noncoding sequence, which is identical to other HRG isoform sequences and includes a polyadenylation signal followed by an A-rich region. The open reading frame encodes a protein of 768 amino acid residues with a calculated molecular mass of 84.2 kDa.

(d) The selection of HRG- β 1 variants containing residues corresponding to the minimal EGF-like domain (HRG- β 1 177-228) was conducted using monovalent phage display. For these variants, residue numbers also are expressed, in parentheses, in terms of the position of the residue in the minimal EGF-like domain (i.e., HRG- β 1 EGF 1-52).

Variants of HRG- β 1 EGF were prepared and selected for binding to HER-3-Ig using monovalent phage display, according to the method of Bass *et al.*, *Proteins* 8:309-314 (1990). As discussed in detail below, an HRG- β 1 EGF phagemid vector was prepared, in which HRG- β 1 EGF was fused to a C-terminal fragment of the M13 coat protein pIII. Kunkel mutagenesis was performed to introduce stop codons into this vector at sites selected for randomization. This step ensures that the starting vector is incapable of expressing the wild-type polypeptide. Stretches of four to six residues per library were randomized in a linear fashion, except for the six cysteines, Phe189 (HRG- β 1 EGF Phe13) and the two most C-terminal residues. Phe189 was not altered because this residue is conserved as an aromatic residue in EGF and TGF- α and forms a stacking interaction with Tyr208 (HRG- β 1 EGF Tyr32) Jacobsen *et al.*, *Biochemistry* 35:3402-

Table 4: Library C Variants

		Position in HRG- β 1				
Variant No.		191	192	193	194	195
Wild-type		V	N	G	G	E
1, 2, 4, 5, 7-12	
3		V
6		Q

Table 5: Library D Variants

		Position in HRG- β 1				
Variant No.		197	198	199	200	201
Wild-type		F	M	V	K	D
1*, 2*, 8*, 12*		Y	K	.	R	I
3		.	R	.	.	T
4, 5, 7, 9		Y	R	.	.	T
6		Y	.	I	.	Y
10		Y	.	.	.	T
11		M	R	.	R	T

*Variant also contained Met226Ile.

Table 6: Library E Variants

		Position in HRG- β 1				
Variant No.		205	206	207	208	209
Wild-type		P	S	R	Y	L
1		T	P	Y	L	M
2, 4		Y	G	Y	L	M
3*		Y	R	Y	R	M
5, 12		T	H	Y	R	G
6		T	H	Y	R	M
7*		Y	K	Y	R	M
8, 9		T	K	Y	R	G
10		Y	K	Y	R	.

*Variant also contained Met226Ile.

Table 7: Library G Variants

		Position in HRG- β 1					
Variant No.		211	212	213	214	215	216
Wild-type		K	C	P	N	E	F
1, 5, 6, 10, 12		R	.	S	L	.	.
2		R	.	S	E	.	.
3		.	.	.	K	.	M
4		R	.	T	V	.	Y
7, 8		R	.	T	V	.	Y
9		.	.	N	S	.	.
11		R	.	K	K	.	.

Example 2 - HER2/HER3 expression in embryonic rat lung

Rat lungs were microdissected from rat embryos on embryologic day (E) 16, 18, 20, and post-natal (P) days 7, 14, and adult. Isolated lung tissue was homogenized in a standard protease inhibitor buffer, and equal protein amounts subjected to SDS-PAGE (4-20%), blotted to nitrocellulose and identified with specific antibodies (HER2, HER3, HER4-Santa Cruz Biological, San Jose, CA. and HRG, 3G11, Genentech, Inc.) using chemiluminescent techniques.

Analysis of the blot indicates that HER2 is expressed at high levels throughout development in utero, appears to peak on E18 and declines after birth to lower adult levels. HER3 expression peaks at E18,

containing overhangs. The fragment was inserted into phagemid display vector pam-g3 by restriction digestion at the same sites to generate construct pHRG2-g3 (177-244). pam-g3 was a derivative of phGHam-g3, which was designed for phage display of human growth hormone (hGH) and was described in Lowman et al., *Biochemistry* 30:10832-10838 (1991). pam-g3 was produced by removing the hGH gene present in phGHam-g3 and replacing this gene with a stuffer fragment, which provides space for cleavage at the restriction sites used for cloning. The HRG- β 1 fragment was attached to residue 247 of pIII.

The HRG- β 1 EGF-like domain expressed from the above-described construct is designated by removing the "p" and the "-g3" that appear in the name of the construct. Thus, the HRG- β 1 EGF-like domain expressed from the pHRG2-g3 construct is designated "HRG2."

The domain was displayed monovalently on phage as a pIII fusion protein, as described by Bass et al., *Proteins* 8:309-314 (1990).

Similarly, variants HRG- β 1₁₄₇₋₂₂₇, HRG- β 1₁₄₇₋₂₄₄, and HRG- β 1₁₇₇₋₂₂₇ were prepared and expressed as described above.

Example 6 - rHRG β 1₁₇₇₋₂₄₄ causes accelerated lung development

Exogenous rHRG β 1₁₇₇₋₂₄₄ caused accelerated lung development *in vitro*. To determine if the expressed HER2/HER3 receptors were functional and the role of HRG stimulation during lung development *in vitro*, rHRG β 1₁₇₇₋₂₄₄ was added to the *in vitro* culture at 10 nM. Tissue was harvested at D5, snap frozen, and 5 micron sections were cut for analysis.

The morphology, in comparison to the untreated control specimens was grossly different. There was marked proliferation of the epithelium. Air spaces which are typically lined with a single cell layer now had a 2-3 cell thickness. The changes were dose dependent with more epithelial cell response with higher concentrations. HER2 and HER3 were still identifiable in the epithelium only.

Example 7 - Human lung differentiation

Differentiation of human lung epithelial cells occurs after HRG treatment. Differentiation was measured by Surfactant Protein A (SPA) production. All sections were stained for SPA. Human lung explant stained for SPA with stain localizing in epithelial cells of the prealveolar ducts. Human lung explant exposed to 10 nM HRG showed an effect on SPA production. As a differentiation control, a lung explant was exposed to 1mM dibutyl cAMP. A negative control was also run.

17. The method of Claim 16, wherein the activating ligand is a heregulin (HRG) polypeptide, HRG variant, HRG agonist antibody or fragment thereof capable of binding to the HER2, HER3 and/or HER4 receptor.

18. A method of treating respiratory distress or emphysema, comprising administering to a patient in need thereof an effective amount of an isolated HER2, HER3 and/or HER4 activating ligand.

19. The method of Claim 18, wherein the activating ligand is a heregulin (HRG) polypeptide, HRG variant, HRG agonist antibody or fragment thereof capable of binding to the HER2, HER3 and/or HER4 receptor.

20. A method, comprising the steps of:

- (a) obtaining a normal epithelial cell sample from a mammal;
- (b) contacting the sample with a ligand which activates HER2, HER3, HER4 or a combination thereof to induce growth and/or proliferation of epithelial cells in the sample and to obtain an expanded sample; and
- (c) re-introducing the expanded sample into the mammal.

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GG	GCG	CGA	GCG	CCT	CAG	CGC	GGC	CGC	TCG	CTC	TCC	CCC	38
	Ala	Arg	Ala	Pro	Gln	Arg	Gly	Arg	Ser	Leu	Ser	Pro	
	1				5					10			
TCG	AGG	GAC	AAA	CTT	TTC	CCA	AAC	CCG	ATC	CGA	GCC	CTT	77
Ser	Arg	Asp	Lys	Leu	Phe	Pro	Asn	Pro	Ile	Arg	Ala	Leu	
		15					20					25	
GGA	CCA	AAC	TCG	CCT	GCG	CCG	AGA	GCC	GTC	CGC	GTA	GAG	116
Gly	Pro	Asn	Ser	Pro	Ala	Pro	Arg	Ala	Val	Arg	Val	Glu	
				30					35				
CGC	TCC	GTC	TCC	GGC	GAG	ATG	TCC	GAG	CGC	AAA	GAA	GGC	155
Arg	Ser	Val	Ser	Gly	Glu	Met	Ser	Glu	Arg	Lys	Glu	Gly	
	40					45					50		
AGA	GGC	AAA	GGG	AAG	GGC	AAG	AAG	AAG	GAG	CGA	GGC	TCC	194
Arg	Gly	Lys	Gly	Lys	Gly	Lys	Lys	Lys	Glu	Arg	Gly	Ser	
			55					60					
GGC	AAG	AAG	CCG	GAG	TCC	GCG	GCG	GGC	AGC	CAG	AGC	CCA	233
Gly	Lys	Lys	Pro	Glu	Ser	Ala	Ala	Gly	Ser	Gln	Ser	Pro	
	65				70					75			
GCC	TTG	CCT	CCC	CGA	TTG	AAA	GAG	ATG	AAA	AGC	CAG	GAA	272
Ala	Leu	Pro	Pro	Arg	Leu	Lys	Glu	Met	Lys	Ser	Gln	Glu	
		80					85					90	
TCG	GCT	GCA	GGT	TCC	AAA	CTA	GTC	CTT	CGG	TGT	GAA	ACC	311
Ser	Ala	Ala	Gly	Ser	Lys	Leu	Val	Leu	Arg	Cys	Glu	Thr	
				95					100				
AGT	TCT	GAA	TAC	TCC	TCT	CTC	AGA	TTC	AAG	TGG	TTC	AAG	350
Ser	Ser	Glu	Tyr	Ser	Ser	Leu	Arg	Phe	Lys	Trp	Phe	Lys	
	105					110					115		
AAT	GGG	AAT	GAA	TTG	AAT	CGA	AAA	AAC	AAA	CCA	CAA	AAT	389
Asn	Gly	Asn	Glu	Leu	Asn	Arg	Lys	Asn	Lys	Pro	Gln	Asn	
			120					125					
ATC	AAG	ATA	CAA	AAA	AAG	CCA	GGG	AAG	TCA	GAA	CTT	CGC	428
Ile	Lys	Ile	Gln	Lys	Lys	Pro	Gly	Lys	Ser	Glu	Leu	Arg	
	130				135					140			
ATT	AAC	AAA	GCA	TCA	CTG	GCT	GAT	TCT	GGA	GAG	TAT	ATG	467
Ile	Asn	Lys	Ala	Ser	Leu	Ala	Asp	Ser	Gly	Glu	Tyr	Met	
		145				150						155	
TGC	AAA	GTG	ATC	AGC	AAA	TTA	GGA	AAT	GAC	AGT	GCC	TCT	506
Cys	Lys	Val	Ile	Ser	Lys	Leu	Gly	Asn	Asp	Ser	Ala	Ser	
				160					165				

FIG. 1A

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GCC	AAT	ATC	ACC	ATC	GTG	GAA	TCA	AAC	GAG	ATC	ATC	ACT	545
Ala	Asn	Ile	Thr	Ile	Val	Glu	Ser	Asn	Glu	Ile	Ile	Thr	
170						175						180	
GGT	ATG	CCA	GCC	TCA	ACT	GAA	GGA	GCA	TAT	GTG	TCT	TCA	584
Gly	Met	Pro	Ala	Ser	Thr	Glu	Gly	Ala	Tyr	Val	Ser	Ser	
			185					190					
GAG	TCT	CCC	ATT	AGA	ATA	TCA	GTA	TCC	ACA	GAA	GGA	GCA	623
Glu	Ser	Pro	Ile	Arg	Ile	Ser	Val	Ser	Thr	Glu	Gly	Ala	
195					200					205			
AAT	ACT	TCT	TCA	TCT	ACA	TCT	ACA	TCC	ACC	ACT	GGG	ACA	662
Asn	Thr	Ser	Ser	Ser	Thr	Ser	Thr	Ser	Thr	Thr	Gly	Thr	
		210					215					220	
AGC	CAT	CTT	GTA	AAA	TGT	GCG	GAG	AAG	GAG	AAA	ACT	TTC	701
Ser	His	Leu	Val	Lys	Cys	Ala	Glu	Lys	Glu	Lys	Thr	Phe	
				225					230				
TGT	GTG	AAT	GGA	GGG	GAG	TGC	TTC	ATG	GTG	AAA	GAC	CTT	740
Cys	Val	Asn	Gly	Gly	Glu	Cys	Phe	Met	Val	Lys	Asp	Leu	
235						240					245		
TCA	AAC	CCC	TCG	AGA	TAC	TTG	TGC	AAG	TGC	CAA	CCT	GGA	779
Ser	Asn	Pro	Ser	Arg	Tyr	Leu	Cys	Lys	Cys	Gln	Pro	Gly	
			250					255					
TTC	ACT	GGA	GCA	AGA	TGT	ACT	GAG	AAT	GTG	CCC	ATG	AAA	818
Phe	Thr	Gly	Ala	Arg	Cys	Thr	Glu	Asn	Val	Pro	Met	Lys	
260					265					270			
GTC	CAA	AAC	CAA	GAA	AAG	GCG	GAG	GAG	CTG	TAC	CAG	AAG	857
Val	Gln	Asn	Gln	Glu	Lys	Ala	Glu	Glu	Leu	Tyr	Gln	Lys	
		275					280					285	
AGA	GTG	CTG	ACC	ATA	ACC	GGC	ATC	TGC	ATC	GCC	CTC	CTT	896
Arg	Val	Leu	Thr	Ile	Thr	Gly	Ile	Cys	Ile	Ala	Leu	Leu	
				290					295				
GTG	GTC	GGC	ATC	ATG	TGT	GTG	GTG	GCC	TAC	TGC	AAA	ACC	935
Val	Val	Gly	Ile	Met	Cys	Val	Val	Ala	Tyr	Cys	Lys	Thr	
300						305					310		
AAG	AAA	CAG	CGG	AAA	AAG	CTG	CAT	GAC	CGT	CTT	CGG	CAG	974
Lys	Lys	Gln	Arg	Lys	Lys	Leu	His	Asp	Arg	Leu	Arg	Gln	
			315					320					
AGC	CTT	CGG	TCT	GAA	CGA	AAC	AAT	ATG	ATG	AAC	ATT	GCC	1013
Ser	Leu	Arg	Ser	Glu	Arg	Asn	Asn	Met	Met	Asn	Ile	Ala	
325					330					335			

FIG. 1B

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AAT	GGG	CCT	CAC	CAT	CCT	AAC	CCA	CCC	CCC	GAG	AAT	GTC	1052
Asn	Gly	Pro	His	His	Pro	Asn	Pro	Pro	Pro	Glu	Asn	Val	
		340					345					350	
CAG	CTG	GTG	AAT	CAA	TAC	GTA	TCT	AAA	AAC	GTC	ATC	TCC	1091
Gln	Leu	Val	Asn	Gln	Tyr	Val	Ser	Lys	Asn	Val	Ile	Ser	
			355						360				
AGT	GAG	CAT	ATT	GTT	GAG	AGA	GAA	GCA	GAG	ACA	TCC	TTT	1130
Ser	Glu	His	Ile	Val	Glu	Arg	Glu	Ala	Glu	Thr	Ser	Phe	
	365					370					375		
TCC	ACC	AGT	CAC	TAT	ACT	TCC	ACA	GCC	CAT	CAC	TCC	ACT	1169
Ser	Thr	Ser	His	Tyr	Thr	Ser	Thr	Ala	His	His	Ser	Thr	
			380					385					
ACT	GTC	ACC	CAG	ACT	CCT	AGC	CAC	AGC	TGG	AGC	AAC	GGA	1208
Thr	Val	Thr	Gln	Thr	Pro	Ser	His	Ser	Trp	Ser	Asn	Gly	
	390				395					400			
CAC	ACT	GAA	AGC	ATC	CTT	TCC	GAA	AGC	CAC	TCT	GTA	ATC	1247
His	Thr	Glu	Ser	Ile	Leu	Ser	Glu	Ser	His	Ser	Val	Ile	
		405					410					415	
GTG	ATG	TCA	TCC	GTA	GAA	AAC	AGT	AGG	CAC	AGC	AGC	CCA	1286
Val	Met	Ser	Ser	Val	Glu	Asn	Ser	Arg	His	Ser	Ser	Pro	
				420				425					
ACT	GGG	GGC	CCA	AGA	GGA	CGT	CTT	AAT	GGC	ACA	GGA	GGC	1325
Thr	Gly	Gly	Pro	Arg	Gly	Arg	Leu	Asn	Gly	Thr	Gly	Gly	
	430					435					440		
CCT	CGT	GAA	TGT	AAC	AGC	TTC	CTC	AGG	CAT	GCC	AGA	GAA	1364
Pro	Arg	Glu	Cys	Asn	Ser	Phe	Leu	Arg	His	Ala	Arg	Glu	
			445					450					
ACC	CCT	GAT	TCC	TAC	CGA	GAC	TCT	CCT	CAT	AGT	GAA	AGG	1403
Thr	Pro	Asp	Ser	Tyr	Arg	Asp	Ser	Pro	His	Ser	Glu	Arg	
					460					465			
TAT	GTG	TCA	GCC	ATG	ACC	ACC	CCG	GCT	CGT	ATG	TCA	CCT	1442
Tyr	Val	Ser	Ala	Met	Thr	Thr	Pro	Ala	Arg	Met	Ser	Pro	
		470					475					480	
GTA	GAT	TTC	CAC	ACG	CCA	AGC	TCC	CCC	AAA	TCG	CCC	CCT	1481
Val	Asp	Phe	His	Thr	Pro	Ser	Ser	Pro	Lys	Ser	Pro	Pro	
				485					490				
TCG	GAA	ATG	TCT	CCA	CCC	GTG	TCC	AGC	ATG	ACG	GTG	TCC	1520
Ser	Glu	Met	S r	Pro	Pro	Val	Ser	Ser	M t	Thr	Val	Ser	
	495					500					505		

FIG. 1C

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ATG	CCT	TCC	ATG	GCG	GTC	AGC	CCC	TTC	ATG	GAA	GAA	GAG	1559
Met	Pro	Ser	Met	Ala	Val	Ser	Pro	Phe	Met	Glu	Glu	Glu	
			510					515					
AGA	CCT	CTA	CTT	CTC	GTG	ACA	CCA	CCA	AGG	CTG	CGG	GAG	1598
Arg	Pro	Leu	Leu	Leu	Val	Thr	Pro	Pro	Arg	Leu	Arg	Glu	
520					525					530			
AAG	AAG	TTT	GAC	CAT	CAC	CCT	CAG	CAG	TTC	AGC	TCC	TTC	1637
Lys	Lys	Phe	Asp	His	His	Pro	Gln	Gln	Phe	Ser	Ser	Phe	
		535					540					545	
CAC	CAC	AAC	CCC	GCG	CAT	GAC	AGT	AAC	AGC	CTC	CCT	GCT	1676
His	His	Asn	Pro	Ala	His	Asp	Ser	Asn	Ser	Leu	Pro	Ala	
				550					555				
AGC	CCC	TTG	AGG	ATA	GTG	GAG	GAT	GAG	GAG	TAT	GAA	ACG	1715
Ser	Pro	Leu	Arg	Ile	Val	Glu	Asp	Glu	Glu	Tyr	Glu	Thr	
	560					565					570		
ACC	CAA	GAG	TAC	GAG	CCA	GCC	CAA	GAG	CCT	GTT	AAG	AAA	1754
Thr	Gln	Glu	Tyr	Glu	Pro	Ala	Gln	Glu	Pro	Val	Lys	Lys	
			575					580					
CTC	GCC	AAT	AGC	CGG	CGG	GCC	AAA	AGA	ACC	AAG	CCC	AAT	1793
Leu	Ala	Asn	Ser	Arg	Arg	Ala	Lys	Arg	Thr	Lys	Pro	Asn	
585					590					595			
GGC	CAC	ATT	GCT	AAC	AGA	TTG	GAA	GTG	GAC	AGC	AAC	ACA	1832
Gly	His	Ile	Ala	Asn	Arg	Leu	Glu	Val	Asp	Ser	Asn	Thr	
		600					605					610	
AGC	TCC	CAG	AGC	AGT	AAC	TCA	GAG	AGT	GAA	ACA	GAA	GAT	1871
Ser	Ser	Gln	Ser	Ser	Asn	Ser	Glu	Ser	Glu	Thr	Glu	Asp	
				615					620				
GAA	AGA	GTA	GGT	GAA	GAT	ACG	CCT	TTC	CTG	GGC	ATA	CAG	1910
Glu	Arg	Val	Gly	Glu	Asp	Thr	Pro	Phe	Leu	Gly	Ile	Gln	
	625					630					635		
AAC	CCC	CTG	GCA	GCC	AGT	CTT	GAG	GCA	ACA	CCT	GCC	TTC	1949
Asn	Pro	Leu	Ala	Ala	Ser	Leu	Glu	Ala	Thr	Pro	Ala	Phe	
			640					645					
CGC	CTG	GCT	GAC	AGC	AGG	ACT	AAC	CCA	GCA	GGC	CGC	TTC	1988
Arg	Leu	Ala	Asp	Ser	Arg	Thr	Asn	Pro	Ala	Gly	Arg	Phe	
650					655					660			
TCG	ACA	CAG	GAA	GAA	ATC	CAG	G						2010
Ser	Thr	Gln	Glu	Glu	Ile	Gln							
		665				669							

FIG. 1D

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GG	GAC	AAA	CTT	TTC	CCA	AAC	CCG	ATC	CGA	GCC	CTT	GGA	38
	Asp	Lys	Leu	Phe	Pro	Asn	Pro	Ile	Arg	Ala	Leu	Gly	
	1				5					10			
CCA	AAC	TCG	CCT	GCG	CCG	AGA	GCC	GTC	CGC	GTA	GAG	CGC	77
Pro	Asn	Ser	Pro	Ala	Pro	Arg	Ala	Val	Arg	Val	Glu	Arg	
		15					20					25	
TCC	GTC	TCC	GGC	GAG	ATG	TCC	GAG	CGC	AAA	GAA	GGC	AGA	116
Ser	Val	Ser	Gly	Glu	Met	Ser	Glu	Arg	Lys	Glu	Gly	Arg	
			30						35				
GGC	AAA	GGG	AAG	GGC	AAG	AAG	AAG	GAG	CGA	GGC	TCC	GGC	155
Gly	Lys	Gly	Lys	Gly	Lys	Lys	Lys	Glu	Arg	Gly	Ser	Gly	
	40					45					50		
AAG	AAG	CCG	GAG	TCC	GCG	GCG	GGC	AGC	CAG	AGC	CCA	GCC	194
Lys	Lys	Pro	Glu	Ser	Ala	Ala	Gly	Ser	Gln	Ser	Pro	Ala	
			55					60					
TTG	CCT	CCC	CAA	TTG	AAA	GAG	ATG	AAA	AGC	CAG	GAA	TCG	233
Leu	Pro	Pro	Gln	Leu	Lys	Glu	Met	Lys	Ser	Gln	Glu	Ser	
65					70					75			
GCT	GCA	GGT	TCC	AAA	CTA	GTC	CTT	CGG	TGT	GAA	ACC	AGT	272
Ala	Ala	Gly	Ser	Lys	Leu	Val	Leu	Arg	Cys	Glu	Thr	Ser	
		80					85					90	
TCT	GAA	TAC	TCC	TCT	CTC	AGA	TTC	AAG	TGG	TTC	AAG	AAT	311
Ser	Glu	Tyr	Ser	Ser	Leu	Arg	Phe	Lys	Trp	Phe	Lys	Asn	
				95					100				
GGG	AAT	GAA	TTG	AAT	CGA	AAA	AAC	AAA	CCA	CAA	AAT	ATC	350
Gly	Asn	Glu	Leu	Asn	Arg	Lys	Asn	Lys	Pro	Gln	Asn	Ile	
	105					110					115		
AAG	ATA	CAA	AAA	AAG	CCA	GGG	AAG	TCA	GAA	CTT	CGC	ATT	389
Lys	Ile	Gln	Lys	Lys	Pro	Gly	Lys	Ser	Glu	Leu	Arg	Ile	
			120					125					
AAC	AAA	GCA	TCA	CTG	GCT	GAT	TCT	GGA	GAG	TAT	ATG	TGC	428
Asn	Lys	Ala	Ser	Leu	Ala	Asp	Ser	Gly	Glu	Tyr	Met	Cys	
130					135					140			
AAA	GTG	ATC	AGC	AAA	TTA	GGA	AAT	GAC	AGT	GCC	TCT	GCC	467
Lys	Val	Ile	Ser	Lys	Leu	Gly	Asn	Asp	Ser	Ala	Ser	Ala	
		145					150					155	
AAT	ATC	ACC	ATC	GTG	GAA	TCA	AAC	GAG	ATC	ATC	ACT	GGT	506
Asn	Ile	Thr	Ile	Val	Glu	Ser	Asn	Glu	Ile	Ile	Thr	Gly	
				160					165				

FIG._2A

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ATG	CCA	GCC	TCA	ACT	GAA	GGA	GCA	TAT	GTG	TCT	TCA	GAG	545
Met	Pro	Ala	Ser	Thr	Glu	Gly	Ala	Tyr	Val	Ser	Ser	Glu	
	170					175					180		
TCT	CCC	ATT	AGA	ATA	TCA	GTA	TCC	ACA	GAA	GGA	GCA	AAT	584
Ser	Pro	Ile	Arg	Ile	Ser	Val	Ser	Thr	Glu	Gly	Ala	Asn	
			185					190					
ACT	TCT	TCA	TCT	ACA	TCT	ACA	TCC	ACC	ACT	GGG	ACA	AGC	623
Thr	Ser	Ser	Ser	Thr	Ser	Thr	Ser	Thr	Thr	Gly	Thr	Ser	
195					200					205			
CAT	CTT	GTA	AAA	TGT	GCG	GAG	AAG	GAG	AAA	ACT	TTC	TGT	662
His	Leu	Val	Lys	Cys	Ala	Glu	Lys	Glu	Lys	Thr	Phe	Cys	
		210					215					220	
GTG	AAT	GGA	GGG	GAG	TGC	TTC	ATG	GTG	AAA	GAC	CTT	TCA	701
Val	Asn	Gly	Gly	Glu	Cys	Phe	Met	Val	Lys	Asp	Leu	Ser	
				225					230				
AAC	CCC	TCG	AGA	TAC	TTG	TGC	AAG	TGC	CCA	AAT	GAG	TTT	740
Asn	Pro	Ser	Arg	Tyr	Leu	Cys	Lys	Cys	Pro	Asn	Glu	Phe	
	235					240					245		
ACT	GGT	GAT	CGC	TGC	CAA	AAC	TAC	GTA	ATG	GCC	AGC	TTC	779
Thr	Gly	Asp	Arg	Cys	Gln	Asn	Tyr	Val	Met	Ala	Ser	Phe	
			250					255					
TAC	AAG	CAT	CTT	GGG	ATT	GAA	TTT	ATG	GAG	GCG	GAG	GAG	818
Tyr	Lys	His	Leu	Gly	Ile	Glu	Phe	Met	Glu	Ala	Glu	Glu	
260					265					270			
CTG	TAC	CAG	AAG	AGA	GTG	CTG	ACC	ATA	ACC	GGC	ATC	TGC	857
Leu	Tyr	Gln	Lys	Arg	Val	Leu	Thr	Ile	Thr	Gly	Ile	Cys	
			275				280					285	
ATC	GCC	CTC	CTT	GTG	GTC	GGC	ATC	ATG	TGT	GTG	GTG	GCC	896
Ile	Ala	Leu	Leu	Val	Val	Gly	Ile	Met	Cys	Val	Val	Ala	
				290					295				
TAC	TGC	AAA	ACC	AAG	AAA	CAG	CGG	AAA	AAG	CTG	CAT	GAC	935
Tyr	Cys	Lys	Thr	Lys	Lys	Gln	Arg	Lys	Lys	Leu	His	Asp	
	300					305					310		
CGT	CTT	CGG	CAG	AGC	CTT	CGG	TCT	GAA	CGA	AAC	AAT	ATG	974
Arg	Leu	Arg	Gln	Ser	Leu	Arg	Ser	Glu	Arg	Asn	Asn	Met	
			315					320					
ATG	AAC	ATT	GCC	AAT	GGG	CCT	CAC	CAT	CCT	AAC	CCA	CCC	1013
Met	Asn	Ile	Ala	Asn	Gly	Pro	His	His	Pro	Asn	Pro	Pro	
325					330					335			

FIG. 2B

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CCC	GAG	AAT	GTC	CAG	CTG	GTG	AAT	CAA	TAC	GTA	TCT	AAA	1052
Pro	Glu	Asn	Val	Gln	Leu	Val	Asn	Gln	Tyr	Val	Ser	Lys	
		340					345					350	
AAC	GTC	ATC	TCC	AGT	GAG	CAT	ATT	GTT	GAG	AGA	GAA	GCA	1091
Asn	Val	Ile	Ser	Ser	Glu	His	Ile	Val	Glu	Arg	Glu	Ala	
				355					360				
GAG	ACA	TCC	TTT	TCC	ACC	AGT	CAC	TAT	ACT	TCC	ACA	GCC	1130
Glu	Thr	Ser	Phe	Ser	Thr	Ser	His	Tyr	Thr	Ser	Thr	Ala	
	365					370					375		
CAT	CAC	TCC	ACT	ACT	GTC	ACC	CAG	ACT	CCT	AGC	CAC	AGC	1169
His	His	Ser	Thr	Thr	Val	Thr	Gln	Thr	Pro	Ser	His	Ser	
			380					385					
TGG	AGC	AAC	GGA	CAC	ACT	GAA	AGC	ATC	CTT	TCC	GAA	AGC	1208
Trp	Ser	Asn	Gly	His	Thr	Glu	Ser	Ile	Leu	Ser	Glu	Ser	
	390					395				400			
CAC	TCT	GTA	ATC	GTG	ATG	TCA	TCC	GTA	GAA	AAC	AGT	AGG	1247
His	Ser	Val	Ile	Val	Met	Ser	Ser	Val	Glu	Asn	Ser	Arg	
		405					410					415	
CAC	AGC	AGC	CCA	ACT	GGG	GGC	CCA	AGA	GGA	CGT	CTT	AAT	1286
His	Ser	Ser	Pro	Thr	Gly	Gly	Pro	Arg	Gly	Arg	Leu	Asn	
				420					425				
GGC	ACA	GGA	GGC	CCT	CGT	GAA	TGT	AAC	AGC	TTC	CTC	AGG	1325
Gly	Thr	Gly	Gly	Pro	Arg	Glu	Cys	Asn	Ser	Phe	Leu	Arg	
	430					435					440		
CAT	GCC	AGA	GAA	ACC	CCT	GAT	TCC	TAC	CGA	GAC	TCT	CCT	1364
His	Ala	Arg	Glu	Thr	Pro	Asp	Ser	Tyr	Arg	Asp	Ser	Pro	
			445					450					
CAT	AGT	GAA	AGG	TAT	GTG	TCA	GCC	ATG	ACC	ACC	CCG	GCT	1403
His	Ser	Glu	Arg	Tyr	Val	Ser	Ala	Met	Thr	Thr	Pro	Ala	
	455				460					465			
CGT	ATG	TCA	CCT	GTA	GAT	TTC	CAC	ACG	CCA	AGC	TCC	CCC	1442
Arg	Met	Ser	Pro	Val	Asp	Phe	His	Thr	Pro	Ser	Ser	Pro	
			470				475					480	
AAA	TCG	CCC	CCT	TCG	GAA	ATG	TCT	CCA	CCC	GTG	TCC	AGC	1481
Lys	Ser	Pro	Pro	Ser	Glu	Met	Ser	Pro	Pro	Val	Ser	Ser	
				485					490				
ATG	ACG	GTG	TCC	ATG	CCT	TCC	ATG	GCG	GTG	AGC	CCC	TTC	1520
Met	Thr	Val	Ser	Met	Pro	Ser	Met	Ala	Val	Ser	Pro	Phe	
	495					500					505		

FIG. 2C

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ATG	GAA	GAA	GAG	AGA	CCT	CTA	CTT	CTC	GTG	ACA	CCA	CCA	1559
Met	Glu	Glu	Glu	Arg	Pro	Leu	Leu	Leu	Val	Thr	Pro	Pro	
			510					515					
AGG	CTG	CGG	GAG	AAG	AAG	TTT	GAC	CAT	CAC	CCT	CAG	CAG	1598
Arg	Leu	Arg	Glu	Lys	Lys	Phe	Asp	His	His	Pro	Gln	Gln	
520					525					530			
TTC	AGC	TCC	TTC	CAC	CAC	AAC	CCC	GCG	CAT	GAC	AGT	AAC	1637
Phe	Ser	Ser	Phe	His	His	Asn	Pro	Ala	His	Asp	Ser	Asn	
		535					540					545	
AGC	CTC	CCT	GCT	AGC	CCC	TTG	AGG	ATA	GTG	GAG	GAT	GAG	1676
Ser	Leu	Pro	Ala	Ser	Pro	Leu	Arg	Ile	Val	Glu	Asp	Glu	
			550						555				
GAG	TAT	GAA	ACG	ACC	CAA	GAG	TAC	GAG	CCA	GCC	CAA	GAG	1715
Glu	Tyr	Glu	Thr	Thr	Gln	Glu	Tyr	Glu	Pro	Ala	Gln	Glu	
	560					565					570		
CCT	GTT	AAG	AAA	CTC	GCC	AAT	AGC	CGG	CGG	GCC	AAA	AGA	1754
Pro	Val	Lys	Lys	Leu	Ala	Asn	Ser	Arg	Arg	Ala	Lys	Arg	
			575					580					
ACC	AAG	CCC	AAT	GGC	CAC	ATT	GCT	AAC	AGA	TTG	GAA	GTG	1793
Thr	Lys	Pro	Asn	Gly	His	Ile	Ala	Asn	Arg	Leu	Glu	Val	
585					590					595			
GAC	AGC	AAC	ACA	AGC	TCC	CAG	AGC	AGT	AAC	TCA	GAG	AGT	1832
Asp	Ser	Asn	Thr	Ser	Ser	Gln	Ser	Ser	Asn	Ser	Glu	Ser	
		600					605					610	

FIG. 2D

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GAA	ACA	GAA	GAT	GAA	AGA	GTA	GGT	GAA	GAT	ACG	CCT	TTC	1871
Glu	Thr	Glu	Asp	Glu	Arg	Val	Gly	Glu	Asp	Thr	Pro	Phe	
				615					620				
CTG	GGC	ATA	CAG	AAC	CCC	CTG	GCA	GCC	AGT	CTT	GAG	GCA	1910
Leu	Gly	Ile	Gln	Asn	Pro	Leu	Ala	Ala	Ser	Leu	Glu	Ala	
	625					630					635		
ACA	CCT	GCC	TTC	CGC	CTG	GCT	GAC	AGC	AGG	ACT	AAC	CCA	1949
Thr	Pro	Ala	Phe	Arg	Leu	Ala	Asp	Ser	Arg	Thr	Asn	Pro	
			640					645					
GCA	GGC	CGC	TTC	TCG	ACA	CAG	GAA	GAA	ATC	CAG	GCC	AGG	1988
Ala	Gly	Arg	Phe	Ser	Thr	Gln	Glu	Glu	Ile	Gln	Ala	Arg	
650					655					660			
CTG	TCT	AGT	GTA	ATT	GCT	AAC	CAA	GAC	CCT	ATT	GCT	GTA	TA 2029
Leu	Ser	Ser	Val	Ile	Ala	Asn	Gln	Asp	Pro	Ile	Ala	Val	
		665					670					675	
A AACCTAAATA AACACATAGA TTCACCTGTA AAACCTTTATT													2070
TTATATAATA AAGTATTCCA CCTTAAATTA AACCAATTTAT TTTATTTTAG													2120
CAGTTCTGCA AATAGAAAAC AGGAAAAAAA CTTTTATAAA TTAAATATAT													2170
GTATGTAAAA ATGAAAAAAA AAAAAAAA													2199

FIG. 2E

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GTGGCTGCGG GGCAATTGAA AAAGAGCCGG CGAGGAGTTC CCCGAAACTT 50
 GTTGGAAGTC CGGGCTCGCG CGGAGGCCAG GAGCTGAGCG GCGGCGGCTG 100
 COGGACGATG GGAGCGTGAG CAGGACGGTG ATAACCTCTC CCCGATCGGG 150
 TTGCGAGGGC GCCGGGCAGA GGCCAGGACG CGAGCCGCCA GCGGCGGGAC 200
 CCATCGACGA CTTCCCGGGG CGACAGGAGC AGCCCCGAGA GCCAGGGCGA 250
 GCGCCCGTTC CAGGTGGCCG GACCGCCCGC CGCGTCCGCG CCGCGCTCCC 300
 TGCAGGCAAC GGGAGACGCC CCCGCGCAGC GCGAGCGCCT CAGCGCGGCC 350
 GCTCGCTCTC CCCATCGAGG GACAACTTT TCCCAAACCC GATCCGAGCC 400
 CTTGGACCAA ACTCGCCTGC GCCGAGAGCC GTCCGCGTAG AGCGCTCCGT 450
 CTCCGGCGAG ATG TCC GAG CGC AAA GAA GGC AGA GGC AAA 490
 Met Ser Glu Arg Lys Glu Gly Arg Gly Lys
 1 5 10
 GGG AAG GGC AAG AAG AAG GAG CGA GGC TCC GGC AAG AAG 529
 Gly Lys Gly Lys Lys Lys Glu Arg Gly Ser Gly Lys Lys
 15 20
 CCG GAG TCC GCG GCG GGC AGC CAG AGC CCA GCC TTG CCT 568
 Pro Glu Ser Ala Ala Gly Ser Gln Ser Pro Ala Leu Pro
 25 30 35
 CCC CAA TTG AAA GAG ATG AAA AGC CAG GAA TCG GCT GCA 607
 Pro Gln Leu Lys Glu Met Lys Ser Gln Glu Ser Ala Ala
 40 45
 GGT TCC AAA CTA GTC CTT CGG TGT GAA ACC AGT TCT GAA 646
 Gly Ser Lys Leu Val Leu Arg Cys Glu Thr Ser Ser Glu
 50 55 60
 TAC TCC TCT CTC AGA TTC AAG TGG TTC AAG AAT GGG AAT 685
 Tyr Ser Ser Leu Arg Ph Lys Trp Phe Lys Asn Gly Asn
 65 70 75
 GAA TTG AAT CGA AAA AAC AAA CCA CAA AAT ATC AAG ATA 724
 Glu Leu Asn Arg Lys Asn Lys Pro Gln Asn Ile Lys Ile
 80 85

FIG. 3A

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CAA	AAA	AAG	CCA	GGG	AAG	TCA	GAA	CTT	CGC	ATT	AAC	AAA	763
Gln	Ly	Lys	Pro	Gly	Lys	S r	Glu	Leu	Arg	Il	Asn	Lys	
	90					95					100		
GCA	TCA	CTG	GCT	GAT	TCT	GGA	GAG	TAT	ATG	TGC	AAA	GTG	802
Ala	Ser	Leu	Ala	Asp	Ser	Gly	Glu	Tyr	Met	Cys	Lys	Val	
			105					110					
ATC	AGC	AAA	TTA	GGA	AAT	GAC	AGT	GCC	TCT	GCC	AAT	ATC	841
Ile	Ser	Lys	Leu	Gly	Asn	Asp	Ser	Ala	Ser	Ala	Asn	Ile	
115					120					125			
ACC	ATC	GTG	GAA	TCA	AAC	GAG	ATC	ATC	ACT	GGT	ATG	CCA	880
Thr	Ile	Val	Glu	Ser	Asn	Glu	Ile	Ile	Thr	Gly	Met	Pro	
		130					135					140	
GCC	TCA	ACT	GAA	GGA	GCA	TAT	GTG	TCT	TCA	GAG	TCT	CCC	919
Ala	Ser	Thr	Glu	Gly	Ala	Tyr	Val	Ser	Ser	Glu	Ser	Pro	
				145					150				
ATT	AGA	ATA	TCA	GTA	TCC	ACA	GAA	GGA	GCA	AAT	ACT	TCT	958
Ile	Arg	Ile	Ser	Val	Ser	Thr	Glu	Gly	Ala	Asn	Thr	Ser	
	155					160					165		
TCA	TCT	ACA	TCT	ACA	TCC	ACC	ACT	GGG	ACA	AGC	CAT	CTT	997
Ser	Ser	Thr	Ser	Thr	Ser	Thr	Thr	Gly	Thr	Ser	His	Leu	
			170					175					
GTA	AAA	TGT	GCG	GAG	AAG	GAG	AAA	ACT	TTC	TGT	GTG	AAT	1036
Val	Lys	Cys	Ala	Glu	Lys	Glu	Lys	Thr	Phe	Cys	Val	Asn	
180					185					190			
GGA	GGG	GAG	TGC	TTC	ATG	GTG	AAA	GAC	CTT	TCA	AAC	CCC	1075
Gly	Gly	Glu	Cys	Phe	Met	Val	Lys	Asp	Leu	Ser	Asn	Pro	
		195					200					205	
TCG	AGA	TAC	TTG	TGC	AAG	TGC	CCA	AAT	GAG	TTT	ACT	GGT	1114
Ser	Arg	Tyr	Leu	Cys	Lys	Cys	Pro	Asn	Glu	Phe	Thr	Gly	
				210					215				
GAT	CGC	TGC	CAA	AAC	TAC	GTA	ATG	GCC	AGC	TTC	TAC	AAG	1153
Asp	Arg	Cys	Gln	Asn	Tyr	Val	Met	Ala	Ser	Phe	Tyr	Lys	
	220					225					230		
GCG	GAG	GAG	CTG	TAC	CAG	AAG	AGA	GTG	CTG	ACC	ATA	ACC	1192
Ala	Glu	Glu	Leu	Tyr	Gln	Lys	Arg	Val	Leu	Thr	Ile	Thr	
			235					240					
GGC	ATC	TGC	ATC	GCC	CTC	CTT	GTG	GTC	GGC	ATC	ATG	TGT	1231
Gly	Ile	Cys	Ile	Ala	Leu	Leu	Val	Val	Gly	Ile	Met	Cys	
245					250					255			
GTG	GTG	GCC	TAC	TGC	AAA	ACC	AAG	AAA	CAG	CGG	AAA	AAG	1270
Val	Val	Ala	Tyr	Cys	Lys	Thr	Lys	Lys	Gln	Arg	Lys	Lys	
		260					265					270	

FIG. 3B

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CTG	CAT	GAC	CGT	CTT	CGG	CAG	AGC	CTT	CGG	TCT	GAA	CGA	1309
Leu	His	Asp	Arg	Leu	Arg	Gln	Ser	Leu	Arg	Ser	Glu	Arg	
				275					280				

AAC	AAT	ATG	ATG	AAC	ATT	GCC	AAT	GGG	CCT	CAC	CAT	CCT	1348
Asn	Asn	Met	Met	Asn	Ile	Ala	Asn	Gly	Pro	His	His	Pro	
	285					290					295		

AAC	CCA	CCC	CCC	GAG	AAT	GTC	CAG	CTG	GTG	AAT	CAA	TAC	1387
Asn	Pro	Pro	Pro	Glu	Asn	Val	Gln	Leu	Val	Asn	Gln	Tyr	
			300					305					

GTA	TCT	AAA	AAC	GTC	ATC	TCC	AGT	GAG	CAT	ATT	GTT	GAG	1426
Val	Ser	Lys	Asn	Val	Ile	Ser	Ser	Glu	His	Ile	Val	Glu	
310					315					320			

AGA	GAA	GCA	GAG	ACA	TCC	TTT	TCC	ACC	AGT	CAC	TAT	ACT	1465
Arg	Glu	Ala	Glu	Thr	Ser	Phe	Ser	Thr	Ser	His	Tyr	Thr	
		325					330					335	

TCC	ACA	GCC	CAT	CAC	TCC	ACT	ACT	GTC	ACC	CAG	ACT	CCT	1504
Ser	Thr	Ala	His	His	Ser	Thr	Thr	Val	Thr	Gln	Thr	Pro	
				340						345			

AGC	CAC	AGC	TGG	AGC	AAC	GGA	CAC	ACT	GAA	AGC	ATC	CTT	1543
Ser	His	Ser	Trp	Ser	Asn	Gly	His	Thr	Glu	Ser	Ile	Leu	
	350					355					360		

TCC	GAA	AGC	CAC	TCT	GTA	ATC	GTG	ATG	TCA	TCC	GTA	GAA	1582
Ser	Glu	Ser	His	Ser	Val	Ile	Val	Met	Ser	Ser	Val	Glu	
			365					370					

AAC	AGT	AGG	CAC	AGC	AGC	CCA	ACT	GGG	GGC	CCA	AGA	GGA	1621
Asn	Ser	Arg	His	Ser	Ser	Pro	Thr	Gly	Gly	Pro	Arg	Gly	
375					380					385			

CGT	CTT	AAT	GGC	ACA	GGA	GGC	CCT	CGT	GAA	TGT	AAC	AGC	1660
Arg	Leu	Asn	Gly	Thr	Gly	Gly	Pro	Arg	Glu	Cys	Asn	Ser	
		390					395					400	

TTC	CTC	AGG	CAT	GCC	AGA	GAA	ACC	CCT	GAT	TCC	TAC	CGA	1699
Phe	Leu	Arg	His	Ala	Arg	Glu	Thr	Pro	Asp	Ser	Tyr	Arg	
				405					410				

GAC	TCT	CCT	CAT	AGT	GAA	AGG	TAT	GTG	TCA	GCC	ATG	ACC	1738
Asp	Ser	Pro	His	Ser	Glu	Arg	Tyr	Val	Ser	Ala	Met	Thr	
	415					420					425		

ACC	CCG	GCT	CGT	ATG	TCA	CCT	GTA	GAT	TTC	CAC	ACG	CCA	1777
Thr	Pr	Ala	Arg	Met	Ser	Pro	Val	Asp	Phe	His	Thr	Pr	
			430					435					

AGC	TCC	CCC	AAA	TCG	CCC	CCT	TCG	GAA	ATG	TCT	CCA	CCC	1816
Ser	Ser	Pr	Lys	Ser	Pro	Pro	Ser	Glu	Met	Ser	Pro	Pro	
440					445					450			

FIG._3C

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GTG	TCC	AGC	ATG	ACG	GTG	TCC	AAG	CCT	TCC	ATG	GCG	GTC	1855
Val	Ser	Ser	Met	Thr	Val	Ser	Lys	Pro	Ser	Met	Ala	Val	
		455					460					465	
AGC	CCC	TTC	ATG	GAA	GAA	GAG	AGA	CCT	CTA	CTT	CTC	GTG	1894
Ser	Pro	Phe	Met	Glu	Glu	Glu	Arg	Pro	Leu	Leu	Leu	Val	
				470					475				
ACA	CCA	CCA	AGG	CTG	CGG	GAG	AAG	AAG	TTT	GAC	CAT	CAC	1933
Thr	Pro	Pro	Arg	Leu	Arg	Glu	Lys	Lys	Phe	Asp	His	His	
		480				485					490		
CCT	CAG	CAG	TTC	AGC	TCC	TTC	CAC	CAC	AAC	CCC	GCG	CAT	1972
Pro	Gln	Gln	Phe	Ser	Ser	Phe	His	His	Asn	Pro	Ala	His	
			495					500					
GAC	AGT	AAC	AGC	CTC	CCT	GCT	AGC	CCC	TTG	AGG	ATA	GTG	2011
Asp	Ser	Asn	Ser	Leu	Pro	Ala	Ser	Pro	Leu	Arg	Ile	Val	
505					510					515			
GAG	GAT	GAG	GAG	TAT	GAA	ACG	ACC	CAA	GAG	TAC	GAG	CCA	2050
Glu	Asp	Glu	Glu	Tyr	Glu	Thr	Thr	Gln	Glu	Tyr	Glu	Pro	
		520					525					530	
GCC	CAA	GAG	CCT	GTT	AAG	AAA	CTC	GCC	AAT	AGC	CGG	CGG	2089
Ala	Gln	Glu	Pro	Val	Lys	Lys	Leu	Ala	Asn	Ser	Arg	Arg	
				535					540				
GCC	AAA	AGA	ACC	AAG	CCC	AAT	GGC	CAC	ATT	GCT	AAC	AGA	2128
Ala	Lys	Arg	Thr	Lys	Pro	Asn	Gly	His	Ile	Ala	Asn	Arg-	
	545					550					555		
TTG	GAA	GTG	GAC	AGC	AAC	ACA	AGC	TCC	CAG	AGC	AGT	AAC	2167
Leu	Glu	Val	Asp	Ser	Asn	Thr	Ser	Ser	Gln	Ser	Ser	Asn	
			560					565					
TCA	GAG	AGT	GAA	ACA	GAA	GAT	GAA	AGA	GTA	GGT	GAA	GAT	2206
Ser	Glu	Ser	Glu	Thr	Glu	Asp	Glu	Arg	Val	Gly	Glu	Asp	
570					575					580			
ACG	CCT	TTC	CTG	GGC	ATA	CAG	AAC	CCC	CTG	GCA	GCC	AGT	2245
Thr	Pro	Phe	Leu	Gly	Ile	Gln	Asn	Pro	Leu	Ala	Ala	Ser	
		585					590					595	
CTT	GAG	GCA	ACA	CCT	GCC	TTC	CGC	CTG	GCT	GAC	AGC	AGG	2284
Leu	Glu	Ala	Thr	Pro	Ala	Phe	Arg	Leu	Ala	Asp	Ser	Arg	
				600					605				
ACT	AAC	CCA	GCA	GGC	CGC	TTC	TCG	ACA	CAG	GAA	GAA	ATC	2323
Thr	Asn	Pro	Ala	Gly	Arg	Ph	Ser	Thr	Gln	Glu	Glu	Ile	
	610					615					620		
CAG	GCC	AGG	CTG	TCT	AGT	GTA	ATT	GCT	AAC	CAA	GAC	CCT	2362
Gln	Ala	Arg	Leu	Ser	Ser	Val	Ile	Ala	Asn	Gln	Asp	Pro	
			625					630					

FIG. 3D

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ATT GCT GTA TAAACCTA AATAACACA TAGATTCACC TGTAAACTT 2410
Ile Ala Val
635 637

TATTTTATAT AATAAGTAT TCCACCTTAA ATTAAACAAT TTATTTTATT 2460

TTAGCAGTTC TGCAAATAAA AAAAAAAAAA 2490

FIG._3E

GCGCCTGCT CCAACCTGCG GGCGGGAGGT GGGTGGCTGC GGGGCAATTG 50

AAAAAGAGCC GGCGAGGAGT TCCCCGAAAC TTGTTGGAAC TCCGGGCTCG 100

CGCGGAGGCC AGGAGCTGAG CGGCGGCGGC TGCCGGACGA TGGGAGCGTG 150

AGCAGGACGG TGATAACCTC TCCCCGATCG GGTTGCGAGG GCGCCGGGCA 200

GAGGCCAGGA CGCGAGCCGC CAGCGGCGGG ACCCATCGAC GACTTCCCGG 250

GGCGACAGGA GCAGCCCCGA GAGCCAGGGC GAGCGCCCGT TCCAGGTGGC 300

CGGACCGCCC GCCGCGTCCG CGCCGCGCTC CCTGCAGGCA ACGGGAGACG 350

CCCCCGCGCA GCGCGAGCGC CTCAGCGCGG CCGCTCGCTC TCCCCATCGA 400

GGGACAAACT TTTCCCAAAC CCGATCCGAG CCCTTGGACC AAActCGCCT 450

GCGCCGAGAG CCGTCCGCGT AGAGCGCTCC GTCTCCGGCG AG ATG 495
Met
1

TCC GAG CGC AAA GAA GGC AGA GGC AAA GGG AAG GGC AAG 534
Ser Glu Arg Lys Glu Gly Arg Gly Lys Gly Lys Gly Lys
5 10

AAG AAG GAG CGA GGC TCC GGC AAG AAG CCG GAG TCC GCG 573
Lys Lys Glu Arg Gly Ser Gly Lys Lys Pro Glu Ser Ala
15 20 25

GCG GGC AGC CAG AGC CCA GCC TTG CCT CCC CAA TTG AAA 612
Ala Gly Ser Gln Ser Pro Ala Leu Pro Pro Gln Leu Lys
30 35 40

GAG ATG AAA AGC CAG GAA TCG GCT GCA GGT TCC AAA CTA 651
Glu Met Lys Ser Gln Glu Ser Ala Ala Gly Ser Lys Leu
45 50

GTC CTT CGG TGT GAA ACC AGT TCT GAA TAC TCC TCT CTC 690
Val Leu Arg Cys Glu Thr Ser Ser Glu Tyr Ser Ser Leu
55 60 65

AGA TTC AAG TGG TTC AAG AAT GGG AAT GAA TTG AAT CGA 729
Arg Phe Lys Trp Phe Lys Asn Gly Asn Glu Leu Asn Arg
70 75

FIG. 4A

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AAA AAC AAA CCA CAA AAT ATC AAG ATA CAA AAA AAG CCA 768
 Lys Asn Lys Pro Gln Asn Ile Lys Ile Gln Lys Lys Pro
 80 85 90

GGA AAG TCA GAA CTT CGC ATT AAC AAA GCA TCA CTG GCT 807
 Gly Lys Ser Glu Leu Arg Ile Asn Lys Ala Ser Leu Ala
 95 100 105

GAT TCT GGA GAG TAT ATG TGC AAA GTG ATC AGC AAA TTA 846
 Asp Ser Gly Glu Tyr Met Cys Lys Val Ile Ser Lys Leu
 110 115

GGA AAT GAC AGT GCC TCT GCC AAT ATC ACC ATC GTG GAA 885
 Gly Asn Asp Ser Ala Ser Ala Asn Ile Thr Ile Val Glu
 120 125 130

TCA AAC GAG ATC ATC ACT GGT ATG CCA GCC TCA ACT GAA 924
 Ser Asn Glu Ile Ile Thr Gly Met Pro Ala Ser Thr Glu
 135 140

GGA GCA TAT GTG TCT TCA GAG TCT CCC ATT AGA ATA TCA 963
 Gly Ala Tyr Val Ser Ser Glu Ser Pro Ile Arg Ile Ser
 145 150 155

GTA TCC ACA GAA GGA GCA AAT ACT TCT TCA TCT ACA TCT 1002
 Val Ser Thr Glu Gly Ala Asn Thr Ser Ser Ser Thr Ser
 160 165 170

ACA TCC ACC ACT GGG ACA AGC CAT CTT GTA AAA TGT GCG 1041
 Thr Ser Thr Thr Gly Thr Ser His Leu Val Lys Cys Ala
 175 180

GAG AAG GAG AAA ACT TTC TGT GTG AAT GGA GGG GAG TGC 1080
 Glu Lys Glu Lys Thr Phe Cys Val Asn Gly Gly Glu Cys
 185 190 195

TTC ATG GTG AAA GAC CTT TCA AAC CCC TCG AGA TAC TTG 1119
 Phe Met Val Lys Asp Leu Ser Asn Pro Ser Arg Tyr Leu
 200 205

TGC AAG TGC CCA AAT GAG TTT ACT GGT GAT CGC TGC CAA 1158
 Cys Lys Cys Pro Asn Glu Phe Thr Gly Asp Arg Cys Gln
 210 215 220

AAC TAC GTA ATG GCC AGC TTC TAC AGT ACG TCC ACT CCC 1197
 Asn Tyr Val Met Ala Ser Phe Tyr Ser Thr Ser Thr Pro
 225 230 235

TTT CTG TCT CTG CCT GAA TAGGA GCATGCTCAG TTGGTGCTGC 1240
 Phe Leu Ser Leu Pro Glu
 240 241

TTTCTTGTTG CTGCATCTCC CCTCAGATTC CACCTAGAGC TAGATGTGTC 1290

FIG. 4B

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TTACCAGATC TAATATTGAC TGCCTCTGCC TGTCGCATGA GAACATTAAC 1340
AAAAGCAATT GTATTACTTC CTCTGTTCGC GACTAGTTGG CTCTGAGATA 1390
CTAATAGGTG TGTGAGGCTC CGGATGTTTC TGGAATTGAT ATTGAATGAT 1440
GTGATACAAA TTGATAGTCA ATATCAAGCA GTGAAATATG ATAATAAAGG 1490
CATTTCAAAG TCTCACTTTT ATTGATAAAA TAAAAATCAT TCTACTGAAC 1540
AGTCCATCTT CTTTATACAA TGACCACATC CTGAAAAGGG TGTGCTAAG 1590
CTGTAACCGA TATGCACTTG AAATGATGGT AAGTTAATTT TGATTCAGAA 1640
TGTGTTATTT GTCACAAATA AACATAATAA AAGGAGTTCA GATGTTTTTC 1690
TTCATTAACC AAAAAAAAAA AAAAA 1715

FIG. 4C

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GAGGCGCCTG CCTCCAACCT GCGGGCGGGA GGTGGGTGGC TCGGGGGCAA 50
 TTGAAAAGA GCGGGCGAGG AGTTCCCCGA AACTTGTTGG AACTCCGGGC 100
 TCGCGCGGAG GCCAGGAGCT GAGCGGCGGC GGCTGCCGGA CGATGGGAGC 150
 GTGAGCAGGA CGGTGATAAC CTCTCCCCGA TCGGGTTGCG AGGGCGCCGG 200
 GCAGAGGCCA GGACGCGAGC CGCCAGCGGC GGGACCCATC GACGACTTCC 250
 CGGGGCGACA GGAGCAGCCC CGAGAGCCAG GGCAGCGGCC CGTTCCAGGT 300
 GGCCGGACCG CCCGCCGCGT CCGCGCCGCG CTCCCTGCAG GCAACGGGAG 350
 ACGCCCCCGC GCAGCGCGAG CGCCTCAGCG CGGCCGCTCG CTCTCCCCAT 400
 CGAGGGACAA ACTTTTCCCA AACCCGATCC GAGCCCTTGG ACCAACTCG 450
 CCTGCGCCGA GAGCCGTCCG CGTAGAGCGC TCCGTCTCCG GCGAG AT 497
 Met
 - 1
 G TCC GAG CGC AAA GAA GGC AGA GGC AAA GGG AAG GGC AAG 537
 Ser Glu Arg Lys Glu Gly Arg Gly Lys Gly Lys Gly Lys
 5 10
 AAG AAG GAG CGA GGC TCC GGC AAG AAG CCG GAG TCC GCG 576
 Lys Lys Glu Arg Gly Ser Gly Lys Lys Pro Glu Ser Ala
 15 20 25
 GCG GGC AGC CAG AGC CCA GCC TTG CCT CCC CAA TTG AAA 615
 Ala Gly Ser Gln Ser Pro Ala Leu Pro Pro Gln Leu Lys
 30 35 40
 GAG ATG AAA AGC CAG GAA TCG GCT GCA GGT TCC AAA CTA 654
 Glu Met Lys Ser Gln Glu Ser Ala Ala Gly Ser Lys Leu
 45 50
 GTC CTT CGG TGT GAA ACC AGT TCT GAA TAC TCC TCT CTC 693
 Val Leu Arg Cys Glu Thr Ser Ser Glu Tyr Ser Ser Leu
 55 60 65
 AGA TTC AAG TGG TTC AAG AAT GGG AAT GAA TTG AAT CGA 732
 Arg Phe Lys Trp Phe Lys Asn Gly Asn Glu Leu Asn Arg
 70 75

FIG. 5A

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AAA	AAC	AAA	CCA	CAA	AAT	ATC	AAG	ATA	CAA	AAA	AAG	CCA	771
Lys	Asn	Lys	Pro	Gln	Asn	Ile	Lys	Ile	Gln	Lys	Lys	Pro	
80					85					90			
GGG	AAG	TCA	GAA	CTT	CGC	ATT	AAC	AAA	GCA	TCA	CTG	GCT	810
Gly	Lys	Ser	Glu	Leu	Arg	Ile	Asn	Lys	Ala	Ser	Leu	Ala	
	95					100					105		
GAT	TCT	GGA	GAG	TAT	ATG	TGC	AAA	GTG	ATC	AGC	AAA	TTA	849
Asp	Ser	Gly	Glu	Tyr	Met	Cys	Lys	Val	Ile	Ser	Lys	Leu	
			110						115				
GGA	AAT	GAC	AGT	GCC	TCT	GCC	AAT	ATC	ACC	ATC	GTG	GAA	888
Gly	Asn	Asp	Ser	Ala	Ser	Ala	Asn	Ile	Thr	Ile	Val	Glu	
	120					125					130		
TCA	AAC	GAG	ATC	ATC	ACT	GGT	ATG	CCA	GCC	TCA	ACT	GAA	927
Ser	Asn	Glu	Ile	Ile	Thr	Gly	Met	Pro	Ala	Ser	Thr	Glu	
			135					140					
GGA	GCA	TAT	GTG	TCT	TCA	GAG	TCT	CCC	ATT	AGA	ATA	TCA	966
Gly	Ala	Tyr	Val	Ser	Ser	Glu	Ser	Pro	Ile	Arg	Ile	Ser	
145					150					155			
GTA	TCC	ACA	GAA	GGA	GCA	AAT	ACT	TCT	TCA	TCT	ACA	TCT	1005
Val	Ser	Thr	Glu	Gly	Ala	Asn	Thr	Ser	Ser	Ser	Thr	Ser	
		160					165					170	
ACA	TCC	ACC	ACT	GGG	ACA	AGC	CAT	CTT	GTA	AAA	TGT	GCG	1044
Thr	Ser	Thr	Thr	Gly	Thr	Ser	His	Leu	Val	Lys	Cys	Ala	
				175					180				
GAG	AAG	GAG	AAA	ACT	TTC	TGT	GTG	AAT	GGA	GGG	GAG	TGC	1083
Glu	Lys	Glu	Lys	Thr	Phe	Cys	Val	Asn	Gly	Gly	Glu	Cys	
	185					190					195		
TTC	ATG	GTG	AAA	GAC	CTT	TCA	AAC	CCC	TCG	AGA	TAC	TTG	1122
Phe	Met	Val	Lys	Asp	Leu	Ser	Asn	Pro	Ser	Arg	Tyr	Leu	
			200					205					
TGC	AAG	TGC	CCA	AAT	GAG	TTT	ACT	GGT	GAT	CGC	TGC	CAA	1161
Cys	Lys	Cys	Pro	Asn	Glu	Phe	Thr	Gly	Asp	Arg	Cys	Gln	
210					215					220			
AAC	TAC	GTA	ATG	GCC	AGC	TTC	TAC	AAG	GCG	GAG	GAG	CTG	1200
Asn	Tyr	Val	Met	Ala	Ser	Phe	Tyr	Lys	Ala	Glu	Glu	Leu	
		225					230					235	
TAC	CAG	AAG	AGA	GTG	CTG	ACC	ATA	ACC	GGC	ATC	TGC	ATC	1239
Tyr	Gln	Lys	Arg	Val	Leu	Thr	Ile	Thr	Gly	Ile	Cys	Ile	
				240					245				
GCC	CTC	CTT	GTG	GTC	GGC	ATC	ATG	TGT	GTG	GTG	GCC	TAC	1278
Ala	Leu	Leu	Val	Val	Gly	Ile	Met	Cys	Val	Val	Ala	Tyr	
	250					255					260		

FIG.-5B

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TGC AAA ACC AAG AAA CAG CGG AAA AAG CTG CAT GAC CGT 1317
 Cys Lys Thr Lys Lys Gln Arg Lys Lys Leu His Asp Arg
 265 270

CTT CGG CAG AGC CTT CGG TCT GAA CGA AAC AAT ATG ATG 1356
 Leu Arg Gln Ser Leu Arg Ser Glu Arg Asn Asn Met Met
 275 280 285

AAC ATT GCC AAT GGG CCT CAC CAT CCT AAC CCA CCC CCC 1395
 Asn Ile Ala Asn Gly Pro His His Pro Asn Pro Pro Pro
 290 295 300

GAG AAT GTC CAG CTG GTG AAT CAA TAC GTA TCT AAA AAC 1434
 Glu Asn Val Gln Leu Val Asn Gln Tyr Val Ser Lys Asn
 305 310

GTC ATC TCC AGT GAG CAT ATT GTT GAG AGA GAA GCA GAG 1473
 Val Ile Ser Ser Glu His Ile Val Glu Arg Glu Ala Glu
 315 320 325

ACA TCC TTT TCC ACC AGT CAC TAT ACT TCC ACA GCC CAT 1512
 Thr Ser Phe Ser Thr Ser His Tyr Thr Ser Thr Ala His
 330 335

CAC TCC ACT ACT GTC ACC CAG ACT CCT AGC CAC AGC TGG 1551
 His Ser Thr Thr Val Thr Gln Thr Pro Ser His Ser Trp
 340 345 350

AGC AAC GGA CAC ACT GAA AGC ATC CTT TCC GAA AGC CAC 1590
 Ser Asn Gly His Thr Glu Ser Ile Leu Ser Glu Ser His
 355 360 365

TCT GTA ATC GTG ATG TCA TCC GTA GAA AAC AGT AGG CAC 1629
 Ser Val Ile Val Met Ser Ser Val Glu Asn Ser Arg His
 370 375

AGC AGC CCA ACT GGG GGC CCA AGA GGA CGT CTT AAT GGC 1668
 Ser Ser Pro Thr Gly Gly Pro Arg Gly Arg Leu Asn Gly
 380 385 390

ACA GGA GGC CCT CGT GAA TGT AAC AGC TTC CTC AGG CAT 1707
 Thr Gly Gly Pro Arg Glu Cys Asn Ser Phe Leu Arg His
 395 400

GCC AGA GAA ACC CCT GAT TCC TAC CGA GAC TCT CCT CAT 1746
 Ala Arg Glu Thr Pro Asp Ser Tyr Arg Asp Ser Pro His
 405 410 415

AGT GAA AGG TAAAA CCGAAGGCAA AGCTACTGCA GAGGAGAAAC 1790
 Ser Glu Arg
 420

FIG. 5C

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TCAGTCAGAG AATCCCTGTG AGCACCTGCG GTCTCACCTC AGGAAATCTA 1840
CTCTAATCAG AATAAGGGGC GGCAGTTACC TGTTC TAGGA GTGCTCCTAG 1890
TTGATGAAGT CATCTCTTTG TTTGACGGAA CTTATTTCTT CTGAGCTTCT 1940
CTCGTCGTCC CAGTGACTGA CAGGCAACAG ACTCTTAAAG AGCTGGGATG 1990
CTTTGATGCG GAAGGTGCAG CACATGGAGT TTCCAGCTCT GGCCATGGGC 2040
TCAGACCCAC TCGGGGTCTC AGTGTCTCA GTTGTAACAT TAGAGAGATG 2090
GCATCAATGC TTGATAAGGA CCCTTCTATA ATTCCAATTG CCAGTTATCC 2140
AAACTCTGAT TCGGTGGTCG AGCTGGCCTC GTGTTCTTAT CTGCTAACCC 2190
TGTCTTACCT TCCAGCCTCA GTTAAGTCAA ATCAAGGGCT ATGTCATTGC 2240
TGAATGTCAT GGGGGGCAAC TGCTTGCCCT CCACCCTATA GTATCTATTT 2290
TATGAAATTC CAAGAAGGGA TGAATAAATA AATCTCTTGG ATGCTGCGTC 2340
TGGCAGTCTT CACGGGTGGT TTCAAAGCA GAAAAAAAAA AAAAAAAAAA 2390
AAAAAAAAA AAAAAAAAAA AAAAAAAAAA AAAAAAAAAA A 2431

FIG._5D

16	1	M	S	E	R	K	E	G	R	G	K	G	K	K	E	R	G	S	G	K	K	P	E	S	A	A	G	S	Q	S	P	A	L	P	P	R	L	K	E	M	K	S	Q	E	S	A	A	G
11	1	M	S	E	R	K	E	G	R	G	K	G	K	K	E	R	G	S	G	K	K	P	E	S	A	A	G	S	Q	S	P	A	L	P	P	Q	L	K	E	M	K	S	Q	E	S	A	A	G
76	1	M	S	E	R	K	E	G	R	G	K	G	K	K	E	R	G	S	G	K	K	P	E	S	A	A	G	S	Q	S	P	A	L	P	P	Q	L	K	E	M	K	S	Q	E	S	A	A	G
84	1	M	S	E	R	K	E	G	R	G	K	G	K	K	E	R	G	S	G	K	K	P	E	S	A	A	G	S	Q	S	P	A	L	P	P	Q	L	K	E	M	K	S	Q	E	S	A	A	G
78	1	M	S	E	R	K	E	G	R	G	K	G	K	K	E	R	G	S	G	K	K	P	E	S	A	A	G	S	Q	S	P	A	L	P	P	Q	L	K	E	M	K	S	Q	E	S	A	A	G

16	51	S	K	L	V	L	R	C	E	T	S	S	E	Y	S	S	L	R	F	K	W	F	K	N	G	N	E	L	N	R	K	N	K	P	Q	N	I	K	I	Q	K	K	P	G	K	S	E	L	R	I	N
11	51	S	K	L	V	L	R	C	E	T	S	S	E	Y	S	S	L	R	F	K	W	F	K	N	G	N	E	L	N	R	K	N	K	P	Q	N	I	K	I	Q	K	K	P	G	K	S	E	L	R	I	N
76	51	S	K	L	V	L	R	C	E	T	S	S	E	Y	S	S	L	R	F	K	W	F	K	N	G	N	E	L	N	R	K	N	K	P	Q	N	I	K	I	Q	K	K	P	G	K	S	E	L	R	I	N
84	51	S	K	L	V	L	R	C	E	T	S	S	E	Y	S	S	L	R	F	K	W	F	K	N	G	N	E	L	N	R	K	N	K	P	Q	N	I	K	I	Q	K	K	P	G	K	S	E	L	R	I	N
78	51	S	K	L	V	L	R	C	E	T	S	S	E	Y	S	S	L	R	F	K	W	F	K	N	G	N	E	L	N	R	K	N	K	P	Q	N	I	K	I	Q	K	K	P	G	K	S	E	L	R	I	N

16	101	K	A	S	L	A	D	S	G	E	Y	M	C	K	V	I	S	K	L	G	N	D	S	A	S	A	N	I	T	I	V	E	S	N	E	I	I	T	G	M	P	A	S	T	E	G	A	Y	V	S	S
11	101	K	A	S	L	A	D	S	G	E	Y	M	C	K	V	I	S	K	L	G	N	D	S	A	S	A	N	I	T	I	V	E	S	N	E	I	I	T	G	M	P	A	S	T	E	G	A	Y	V	S	S
76	101	K	A	S	L	A	D	S	G	E	Y	M	C	K	V	I	S	K	L	G	N	D	S	A	S	A	N	I	T	I	V	E	S	N	E	I	I	T	G	M	P	A	S	T	E	G	A	Y	V	S	S
84	101	K	A	S	L	A	D	S	G	E	Y	M	C	K	V	I	S	K	L	G	N	D	S	A	S	A	N	I	T	I	V	E	S	N	E	I	I	T	G	M	P	A	S	T	E	G	A	Y	V	S	S
78	101	K	A	S	L	A	D	S	G	E	Y	M	C	K	V	I	S	K	L	G	N	D	S	A	S	A	N	I	T	I	V	E	S	N	E	I	I	T	G	M	P	A	S	T	E	G	A	Y	V	S	S

16	151	E	S	P	I	R	I	S	V	S	T	E	G	A	N	T	S	S	S	T	S	T	T	G	T	S	H	L	V	K	C	A	E	K	E	K	T	F	C	V	N	G	G	E	C	F	M	V	K
11	151	E	S	P	I	R	I	S	V	S	T	E	G	A	N	T	S	S	S	T	S	T	T	G	T	S	H	L	V	K	C	A	E	K	E	K	T	F	C	V	N	G	G	E	C	F	M	V	K
76	151	E	S	P	I	R	I	S	V	S	T	E	G	A	N	T	S	S	S	T	S	T	T	G	T	S	H	L	V	K	C	A	E	K	E	K	T	F	C	V	N	G	G	E	C	F	M	V	K
84	151	E	S	P	I	R	I	S	V	S	T	E	G	A	N	T	S	S	S	T	S	T	T	G	T	S	H	L	V	K	C	A	E	K	E	K	T	F	C	V	N	G	G	E	C	F	M	V	K
78	151	E	S	P	I	R	I	S	V	S	T	E	G	A	N	T	S	S	S	T	S	T	T	G	T	S	H	L	V	K	C	A	E	K	E	K	T	F	C	V	N	G	G	E	C	F	M	V	K

FIG.-6A

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16	201	DL	SN	PS	RY	L	CK	C	Q	P	G	F	T	G	A	R	C	T	E	N	V	P	M	K	V	Q	N	Q	-	-	-	-	-	E	K	A	E	E	L	Y	Q	K	R	V	L	T				
11	201	DL	SN	PS	RY	L	CK	C	P	N	E	F	T	G	D	R	C	Q	N	Y	V	M	A	S	F	Y	K	H	L	G	I	E	F	M	E	A	E	E	L	Y	Q	K	R	V	L	T				
76	201	DL	SN	PS	RY	L	CK	C	P	N	E	F	T	G	D	R	C	Q	N	Y	V	M	A	S	F	Y	K	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-			
84	201	DL	SN	PS	RY	L	CK	C	P	N	E	F	T	G	D	R	C	Q	N	Y	V	M	A	S	F	Y	K	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-				
78	201	DL	SN	PS	RY	L	CK	C	P	N	E	F	T	G	D	R	C	Q	N	Y	V	M	A	S	F	Y	K	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-				
16	248	IT	G	I	C	I	A	L	L	V	V	G	I	M	C	V	V	A	Y	C	K	T	K	Q	R	K	K	L	H	D	R	L	R	Q	S	L	R	S	E	R	N	N	M	M	N	I	A	N	G	
11	251	IT	G	I	C	I	A	L	L	V	V	G	I	M	C	V	V	A	Y	C	K	T	K	Q	R	K	K	L	H	D	R	L	R	Q	S	L	R	S	E	R	N	N	M	M	N	I	A	N	G	
76	243	IT	G	I	C	I	A	L	L	V	V	G	I	M	C	V	V	A	Y	C	K	T	K	Q	R	K	K	L	H	D	R	L	R	Q	S	L	R	S	E	R	N	N	M	M	N	I	A	N	G	
84	243	IT	G	I	C	I	A	L	L	V	V	G	I	M	C	V	V	A	Y	C	K	T	K	Q	R	K	K	L	H	D	R	L	R	Q	S	L	R	S	E	R	N	N	M	M	N	I	A	N	G	
16	298	P	H	H	P	N	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P					
11	301	P	H	H	P	N	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P					
76	293	P	H	H	P	N	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P					
84	293	P	H	H	P	N	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P	P					
16	348	T	V	T	Q	T	P	S	H	S	W	S	N	G	H	T	E	S	I	L	S	E	S	H	S	V	I	V	M	S	S	V	E	N	S	R	H	S	S	P	T	G	Q	P	R	Q	R	L	N	G
11	351	T	V	T	Q	T	P	S	H	S	W	S	N	G	H	T	E	S	I	L	S	E	S	H	S	V	I	V	M	S	S	V	E	N	S	R	H	S	S	P	T	G	Q	P	R	Q	R	L	N	G
76	343	T	V	T	Q	T	P	S	H	S	W	S	N	G	H	T	E	S	I	L	S	E	S	H	S	V	I	V	M	S	S	V	E	N	S	R	H	S	S	P	T	G	Q	P	R	Q	R	L	N	G
84	343	T	V	T	Q	T	P	S	H	S	W	S	N	G	H	T	E	S	I	L	S	E	S	H	S	V	I	V	M	S	S	V	E	N	S	R	H	S	S	P	T	G	Q	P	R	Q	R	L	N	G

FIG. 6B

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16	398	GGPRECN	FLRHARE	TPDSYRD	SPHSERY	VSAMTT	PARMS	PVDFHT	PSSSP
11	401	GGPRECN	FLRHARE	TPDSYRD	SPHSERY	VSAMTT	PARMS	PVDFHT	PSSSP
76	393	GGPRECN	FLRHARE	TPDSYRD	SPHSERY	VSAMTT	PARMS	PVDFHT	PSSSP
84	393	GGPRECN	FLRHARE	TPDSYRD	SPHSER
16	446	KSPPS	EMSP	PPVSS	MTVSM	PSMA	VSPFM	EEER	PLLLVT
11	451	KSPPS	EM8P	PPVSS	MTVSM	PSMA	VSPFM	EEER	PLLLVT
76	443	KSPPS	EMSP	PPVSS	MTVSK	PSMA	VSPFM	EEER	PLLLVT
16	498	QFSS	FHHN	PAHDS	NSLP	ASPL	RIVED	EYETT	QEYEP
11	501	QFSS	FHHN	PAHDS	NSLP	ASPL	RIVED	EYETT	QEYEP
76	493	QFSS	FHHN	PAHDS	NSLP	ASPL	RIVED	EYETT	QEYEP
16	548	RAKRT	KPNQ	GHIAN	RLE	VDSNT	SSQSS	NSESE	TEDERV
11	551	RAKRT	KPNQ	GHIAN	RLE	VDSNT	SSQSS	NSESE	TEDERV
76	543	RAKRT	KPNQ	GHIAN	RLE	VDSNT	SSQSS	NSESE	TEDERV
16	598	AASLE	ATPA	FRLA	DSRT	NPAG	RFS	TQEE	IQ.....
11	601	AASLE	ATPA	FRLA	DSRT	NPAG	RFS	TQEE	IQARL
76	593	AASLE	ATPA	FRLA	DSRT	NPAG	RFS	TQEE	IQARL

FIG.-6C

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1 GGGTACCATGGGTCTGGTGAGCGCGTTCCTCCGCCTGAGCGCAACTAGCGGC
51 GGGTCGTGGGCACCTCCAGAAAAGATCCCGCACCATCCTCCAGGATCCAA
101 TGGCCTTGGAGAGAGGGCTGCAGGGCCCACGGACATTGCTGACTCTTCAG
151 AACGTGCTGACATGGAGCCAGGTAGACTGAAATTATCATGTGTCCAAATT
201 AAAATTGCATACTTCAAGGATTATTTGAAGGACTATTCTTAGACCCTTTT
251 AAGAAGATTTAAAGAAAAACCACTCGGCCCTGAGTGCGGCGAGGACCCTG
301 TTTGTGGATGTGGAGGAGCGCGGGCCGGAGGCCATGGACGTGAAGGAGAG
1 M D V K E R
351 GAAGCCTTACCGCTCGCTGACCCGGCGCCGCGACGCCGAGCGCCGCTACA
7 K P Y R S L T R R R D A E R R Y T
401 CCAGCTCGTCCGCGGACAGCGAGGAGGGCAAAGCCCCGCAGAAATCGTAC
24 S S S A D S E E G K A P Q K S Y
451 AGCTCCAGCGAGACCCTGAAGGCCTACGACCAGGACGCCCGCCTAGCCTA
40 S S S E T L K A Y D Q D A R L A Y
501 TGGCAGCCGCGTCAAGGACATTGTGCCGACAGGAGGCCGAGGAATTCTGCC
57 G S R V K D I V P Q E A E E F C R
551 GCACAGGTGCCAACTTCACCCTGCGGGAGCTGGGGCTGGAAGAAGTAACG
74 T G A N F T L R E L G L E E V T
601 CCCCCTCACGGGACCCTGTACCGGACAGACATTGGCCTCCCCCACTGCGG
90 P P H G T L Y R T D I G L P H C G
651 CTA CTCCATGGGGGCTGGCTCTGATGCCGACATGGAGGCTGACACGGTGC
107 Y S M G A G S D A D M E A D T V L
701 TGTCCCCTGAGCACCCCGTGCGTCTGTGGGGCCGGAGCACACGGTCAGGG
124 S P E H P V R L W G R S T R S G
751 CGCAGCTCCTGCCTGTCCAGCCGGGCCAATTCCAATCTCACACTCACCGA
140 R S S C L S S R A N S N L T L T D
801 CACCGAGCATGAAAACACTGAGACTGATCATCCGGGCGGCCTGCAGAACC
157 T E H E N T E T D H P G G L Q N H
851 ACGCGCGGCTCCGGACGCCGCGCCGCGCTCTCGCACGCCCACACCCCC
174 A R L R T P P P P L S H A H T P
901 AACCAGCACCACGCGGCCTCCATTAACCTCCCTGAACCGGGGCAACTTCAC
190 N Q H H A A S I N S L N R G N F T
951 GCCGAGGAGCAACCCCAGCCCGGCCCCCACGGACCACTCGCTCTCCGGAG
207 P R S N P S P A P T D H S L S G E
1001 AGCCCCCTGCCGGCGGCGCCAGGAGCCTGCCACGCCCAGGAGAACTGG
224 P P A G G A Q E P A H A Q E N W
1051 CTGCTCAACAGCAACATCCCCCTGGAGACCAGAAACCTAGGCAAGCAGCC
240 L L N S N I P L E T R N L G K Q P

FIG. 7A

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1101 ATTCCTAGGGACATTGCAGGACAACCTCATTGAGATGGACATTCTCGGCG
257 F L G T L Q D N L I E M D I L G A

1151 CCTCCCGCCATGATGGGGCTTACAGTGACGGGCACTTCCTCTTCAAGCCT
274 S R H D G A Y S D G H F L F K P

1201 GGAGGCACCTCCCCGCTCTTCTGCACCACATCACCAGGGTACCCACTGAC
290 G G T S P L F C T T S P G Y P L T

1251 GTCCAGCACAGTGTACTCTCCTCCGCCCCGACCCCTGCCCCGCAGCACCT
307 S S T V Y S P P P R P L P R S T F

1301 TCGCCCGGCCGGCCTTTAACCTCAAGAAGCCCTCCAAGTACTGTAAGTGG
324 A R P A F N L K K P S K Y C N W

1351 AAGTGCGCAGCCCTGAGCGCCATCGTCATCTCAGCCACTCTGGTCATCCT
340 K C A A L S A I V I S A T L V I L

1401 GCTGGCATACTTTGTGGCCATGCACCTGTTTGGCCTAAACTGGCACCTGC
357 L A Y F V A M H L F G L N W H L Q

1451 AGCCGATGGAGGGGCAGATGTATGAGATCACGGAGGACACAGCCAGCAGT
374 P M E G Q M Y E I T E D T A S S

1501 TGGCCTGTGCCAACCGACGTCTCCCTATAACCCCTCAGGGGGCACTGGCTT
390 W P V P T D V S L Y P S G G T G L

1551 AGAGACCCCTGACAGGAAAGGCAAAGGAACACAGAAGGAAAGCCCAGTA
407 E T P D R K G K G T T E G K P S S

1601 GTTTCCTTTCCAGAGGACAGTTTCATAGATTCTGGAGAAATTGATGTGGGA
424 F F P E D S F I D S G E I D V G

1651 AGGCGAGCTTCCCAGAAGATTCCTCCTGGCACTTTCTGGAGATCTCAAGT
440 R R A S Q K I P P G T F W R S Q V

1701 GTTCATAGACCATCCTGTGCATCTGAAATTCAATGTGTCTCTGGGAAAGG
457 F I D H P V H L K F N V S L G K A

1751 CAGCCCTGGTTGGCATTATGGCAGAAAAGGCCTCCCTCCTTCACATACA
474 A L V G I Y G R K G L P P S H T

1801 CAGTTTGACTTTGTGGAGCTGCTGGATGGCAGGAGGCTCCTAACCCAGGA
490 Q F D F V E L L D G R R L L T Q E

1851 GGCGCGGAGCCTAGAGGGGACCCCGCGCCAGTCTCGGGGAACTGTGCCCC
507 A R S L E G T P R Q S R G T V P P

1901 CCTCCAGCCATGAGACAGGCTTCATCCAGTATTTGGATTCAGGAATCTGG
524 S S H E T G F I Q Y L D S G I W

1951 CACTTGGCTTTTTTACAATGACGGAAAGGAGTCAGAAGTGGTTTCCTTTCT
540 H L A F Y N D G K E S E V V S F L

FIG. 7B

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2001 CACCACTGCCATTGCCTTGCCTCCCCGATTGAAAGAGATGAAAAGCCAGG
557 T T A I A L P P R L K E M K S Q E

2051 AATCGGCTGCAGGTTCCAAACTAGTCCTTCGGTGTGAAACCAGTTCTGAA
574 S A A G S K L V L R C E T S S E

2101 TACTCCTCTCTCAGATTCAAGTGGTTCAAGAATGGGAATGAATTGAATCG
590 Y S S L R F K W F K N G N E L N R

2151 AAAAAACAAACCACAAAATATCAAGATACAAAAAAGCCAGGGAAGTCAG
607 K N K P Q N I K I Q K K P G K S E

2201 AACTTCGCATTAACAAAGCATCACTGGCTGATTCTGGAGAGTATATGTGC
624 L R I N K A S L A D S G E Y M C

2251 AAAGTGATCAGCAAATTAGGAAATGACAGTGCCTCTGCCAATATCACCAT
640 K V I S K L G N D S A S A N I T I

2301 CGTGGAAATCAAACGAGATCATCACTGGTATGCCAGCCTCAACTGAAGGAG
657 V E S N E I I T G M P A S T E G A

2351 CATATGTGTCTTCAGAGTCTCCCATAGAAATATCAGTATCCACAGAAGGA
674 Y V S S E S P I R I S V S T E G

2401 GCAAATACTTCTTCATCTACATCTACATCCACCCTGGGACAAGCCATCT
690 A N T S S S T S T S T T G T S H L

2451 TGTAATAATGTGCGGAGAAGGAGAAAACCTTCTGTGTGAATGGAGGGGAGT
707 V K (C) A E K E K T F (C) V N G G E (C)

2501 GCTTCATGGTGAAAGACCTTTCAAACCCCTCGAGATACTTGTGCAAGTGC
724 F M V K D L S N P S R Y L (C) K (C)

2551 CCAAATGAGTTTACTGGTGATCGCTGCCAAACTACGTAATGGCCAGCTT
740 P N E F T G D R (C) Q N Y V M A S F

2601 CTACAGTACGTCCACTCCCTTTCTGTCTCTGCCTGAATAGGAGCATGCTC
757 Y S T S T P F L S L P E

2651 AGTTGGTGCTGCTTTCTTGTTGCTGCATCTCCCCTCAGATTCCACCTAGA

2701 GCTAGATGTGTCTTACCAGATCTAATATTGACTGCCTCTGCCTGTGCGCAT
2751 GAGAACATTAACAAAAGCAATTGTATTACTTCCTCTGTTCGCGACTAGTT
2801 GGCTCTGAGATACTAATAAGGTGTGTGAGGCTCCGGATGTTTCTGGAATTG
2851 ATATTGAATGATGTGATACAAATTGATAGTCAATATCAAGCAGTGAAATA
2901 TGATAATAAAGGCATTTCAAAGTCTCACTTTTATTGATAAAATAAAAATC
2951 ATTCTACTGAACAGTCCATCTTCTTTATACAAATGACCACATCCTGAAAAG
3001 GGTGTTGCTAAGCTGTAACCGATATGCACTTGAAATGATGGTAAGTTAAT
3051 TTTGATTTCAGAATGTGTTATTTGTACAAATAAACATAATAAAAGGAAAA
3101 AAAAAAAAAA

FIG. 7C

SUBSTITUTE SHEET (RULE 26)

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EGF
-like

976 TGCAAGGTGACAAGGCTGTGTCTCCTTTGAACCATCAGCGGCACCGACACCGAAGAAATCGTATTTTTCCTTTT
 158 Q G D K A V V S F E P S A A P T P K N R I F A F S
 1051 CTTTCTTCCCGTCCACTGCGCCATCCTTCCCTTACCCACCCGGAAACCCTGAGGTGAGAACGCCCAAGTCAGCAA
 183 F L P S T A P S F P S P T R N P E V R T P K S A T
 1126 CTCAGCCACAAACACAGAAACTAATCTCCAAACTGCTCCTNAACTTTTCTACATCTACATCCACTCGGACAA
 208 Q P Q T T E T N L Q T A P K L S T S T S T T G T S
 1201 GCCATCTTGTAATAATGTGGGAGAGAGAGAAACTTTCTGTGTGAATGGAGGGGAGTGCTTTCATGGTGAAGACC
 233 H L V K [C] A E K E K T F [C] V N G G E [C] F H V K D L
 1276 TTTCAAACCCCTCGAGATACCTTGTGCAAGTGCCCAATGAGTTTACTGGTGATCGCTGCCAAACTAGGTAATGG
 258 S N P S R Y L [C] K [C] P N E F T G D R [C] Q N Y V H A
 1351 CCAGCTTCTACAGTACGTCACCTCCCTTCTGTCTGCTGCTGAATAGGAGCATGCTCAGTTGGTGCTGCTTCTT
 283 S F Y S T S T P F L S L P E O
 1426 GTTGTGCAATCTCCCTCAGATTCCACCTAGAGCTAGATGTGTCTTACCAGATCTAATATTGACTGCCCTCTGCCT
 1501 GTGGCATGAGAACAATTAAAGCAATTGTATTACTTCTCTGTTCGGGACTAGTTGGCTCTGAGATACTAATA
 1576 GGTGTGTGAGGCTCCGGATGTTTCTCGAATTGATATTGAATGATGTGATACAAATTGATAGTCMAATATCAAGCAG
 1651 TGAATATGATATAAAGGCATTTCAAAGTCTCACTTTTATTGATAAAATAAAATCATTTCTACTGAACAGTCCA
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FIG.-8B

Sequence Listing

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<120> Use of Heregulin as a Growth Factor

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<150> US 09/020,598

<151> 1998-02-04

<160> 14

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10 <211> 669

<212> PRT

<213> Homo sapiens

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	Pro	Gly	Lys	Ser	Glu	Leu	Arg	Ile	Asn	Lys	Ala	Ser	Leu	Ala	Asp
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	Ser	Gly	Glu	Tyr	Met	Cys	Lys	Val	Ile	Ser	Lys	Leu	Gly	Asn	Asp
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					Cys Ala Glu Lys Glu Lys Thr Phe Cys Val Asn Gly Gly Glu Cys										
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10					Phe Met Val Lys Asp Leu Ser Asn Pro Ser Arg Tyr Leu Cys Lys										
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					Arg Val Leu Thr Ile Thr Gly Ile Cys Ile Ala Leu Leu Val Val										
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					Gly Ile Met Cys Val Val Ala Tyr Cys Lys Thr Lys Lys Gln Arg										
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20					Lys Lys Leu His Asp Arg Leu Arg Gln Ser Leu Arg Ser Glu Arg										
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					Asn Asn Met Met Asn Ile Ala Asn Gly Pro His His Pro Asn Pro										
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25					Pro Pro Glu Asn Val Gln Leu Val Asn Gln Tyr Val Ser Lys Asn										
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					Val Ile Ser Ser Glu His Ile Val Glu Arg Glu Ala Glu Thr Ser										
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					Phe Ser Thr Ser His Tyr Thr Ser Thr Ala His His Ser Thr Thr										
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	tcg agg gac aaa ctt ttc cca aac ccg atc cga gcc ctt 77															

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5	Gly	Pro	Asn	Ser	Pro	Ala	Pro	Arg	Ala	Val	Arg	Val	Glu	
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	Arg	Ser	Val	Ser	Gly	Glu	Met	Ser	Glu	Arg	Lys	Glu	Gly	
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	Gly	Lys	Lys	Pro	Glu	Ser	Ala	Ala	Gly	Ser	Gln	Ser	Pro	
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	Ala	Leu	Pro	Pro	Arg	Leu	Lys	Glu	Met	Lys	Ser	Gln	Glu	
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	tcg	gct	gca	ggt	tcc	aaa	cta	gtc	ctt	cgg	tgt	gaa	acc	311
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	Ser	Ser	Glu	Tyr	Ser	Ser	Leu	Arg	Phe	Lys	Trp	Phe	Lys	
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	atc	aag	ata	caa	aaa	aag	cca	ggg	aag	tca	gaa	ctt	cgc	428
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	Ile	Asn	Lys	Ala	Ser	Leu	Ala	Asp	Ser	Gly	Glu	Tyr	Met	
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	tgc	aaa	gtg	atc	agc	aaa	tta	gga	aat	gac	agt	gcc	tct	506
	Cys	Lys	Val	Ile	Ser	Lys	Leu	Gly	Asn	Asp	Ser	Ala	Ser	
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	gcc	aat	atc	acc	atc	gtg	gaa	tca	aac	gag	atc	atc	act	545
	Ala	Asn	Ile	Thr	Ile	Val	Glu	Ser	Asn	Glu	Ile	Ile	Thr	
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	ggt	atg	cca	gcc	tca	act	gaa	gga	gca	tat	gtg	tct	tca	584
40	Gly	Met	Pro	Ala	Ser	Thr	Glu	Gly	Ala	Tyr	Val	Ser	Ser	
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	Ser Asn Pro Ser Arg Tyr Leu Cys Lys Cys Gln Pro Gly	
	250	255
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	Phe Thr Gly Ala Arg Cys Thr Glu Asn Val Pro Met Lys	
	260	265 270
	gtc caa aac caa gaa aag gcg gag gag ctg tac cag aag	857
20	Val Gln Asn Gln Glu Lys Ala Glu Glu Leu Tyr Gln Lys	
	275	280 285
	aga gtg ctg acc ata acc ggc atc tgc atc gcc ctc ctt	896
	Arg Val Leu Thr Ile Thr Gly Ile Cys Ile Ala Leu Leu	
	290	295
25	gtg gtc ggc atc atg tgt gtg gtg gcc tac tgc aaa acc	935
	Val Val Gly Ile Met Cys Val Val Ala Tyr Cys Lys Thr	
	300	305 310
	aag aaa cag cgg aaa aag ctg cat gac cgt ctt cgg cag	974
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	Ser Leu Arg Ser Glu Arg Asn Asn Met Met Asn Ile Ala	
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	cag ctg gtg aat caa tac gta tct aaa aac gtc atc tcc	1091
	Gln Leu Val Asn Gln Tyr Val Ser Lys Asn Val Ile Ser	
	355	360
40	agt gag cat att gtt gag aga gaa gca gag aca tcc ttt	1130
	Ser Glu His Ile Val Glu Arg Glu Ala Glu Thr Ser Phe	
	365	370 375
	tcc acc agt cac tat act tcc aca gcc cat cac tcc act	1169

	Ser Thr Ser His Tyr Thr Ser Thr Ala His His Ser Thr	
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	act gtc acc cag act cct agc cac agc tgg agc aac gga	1208
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	cac act gaa agc atc ctt tcc gaa agc cac tct gta atc	1247
	His Thr Glu Ser Ile Leu Ser Glu Ser His Ser Val Ile	
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	gtg atg tca tcc gta gaa aac agt agg cac agc agc cca	1286
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	act ggg ggc cca aga gga cgt ctt aat ggc aca gga ggc	1325
	Thr Gly Gly Pro Arg Gly Arg Leu Asn Gly Thr Gly Gly	
	430	435 440
15	cct cgt gaa tgt aac agc ttc ctc agg cat gcc aga gaa	1364
	Pro Arg Glu Cys Asn Ser Phe Leu Arg His Ala Arg Glu	
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	acc cct gat tcc tac cga gac tct cct cat agt gaa agg	1403
	Thr Pro Asp Ser Tyr Arg Asp Ser Pro His Ser Glu Arg	
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	tat gtg tca gcc atg acc acc ccg gct cgt atg tca cct	1442
	Tyr Val Ser Ala Met Thr Thr Pro Ala Arg Met Ser Pro	
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 615 620
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 Arg Val Gly Glu Asp Thr Pro Phe Leu Gly Ile Gln Asn Pro Leu
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 Pro Asn Ser Pro Ala Pro Arg Ala Val Arg Val Glu Arg
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 Ser Val Ser Gly Glu Met Ser Glu Arg Lys Glu Gly Arg
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 25 ggc aaa ggg aag ggc aag aag aag gag cga gcc tcc ggc 155
 Gly Lys Gly Lys Gly Lys Lys Lys Glu Arg Gly Ser Gly
 40 45 50
 aag aag ccg gag tcc gcg gcg ggc agc cag agc cca gcc 194
 Lys Lys Pro Glu Ser Ala Ala Gly Ser Gln Ser Pro Ala
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 Leu Pro Pro Gln Leu Lys Glu Met Lys Ser Gln Glu Ser
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 Ser Glu Tyr Ser Ser Leu Arg Phe Lys Trp Phe Lys Asn
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 Gly Asn Glu Leu Asn Arg Lys Asn Lys Pro Gln Asn Ile
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 Lys Ile Gln Lys Lys Pro Gly Lys Ser Glu Leu Arg Ile
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5 aac aaa gca tca ctg gct gat tct gga gag tat atg tgc 428
 Asn Lys Ala Ser Leu Ala Asp Ser Gly Glu Tyr Met Cys
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 Ser Pro Ile Arg Ile Ser Val Ser Thr Glu Gly Ala Asn
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20 act tct tca tct aca tct aca tcc acc act ggg aca agc 623
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 His Leu Val Lys Cys Ala Glu Lys Glu Lys Thr Phe Cys
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25 gtg aat gga ggg gag tgc ttc atg gtg aaa gac ctt tca 701
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 Tyr Lys His Leu Gly Ile Glu Phe Met Glu Ala Glu Glu
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	Arg Leu Arg Gln Ser Leu Arg Ser Glu Arg Asn Asn Met	
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	Met Asn Ile Ala Asn Gly Pro His His Pro Asn Pro Pro	
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	Glu Thr Ser Phe Ser Thr Ser His Tyr Thr Ser Thr Ala	
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	Gly Thr Gly Gly Pro Arg Glu Cys Asn Ser Phe Leu Arg	
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	Arg Met Ser Pro Val Asp Phe His Thr Pro Ser Ser Pro	
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5 Met Thr Val Ser Met Pro Ser Met Ala Val Ser Pro Phe
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Arg Leu Arg Glu Lys Lys Phe Asp His His Pro Gln Gln
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15 Phe Ser Ser Phe His His Asn Pro Ala His Asp Ser Asn
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Glu Tyr Glu Thr Thr Gln Glu Tyr Glu Pro Ala Gln Glu
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575 580

25 acc aag ccc aat ggc cac att gct aac aga ttg gaa gtg 1793
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585 590 595

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30 Asp Ser Asn Thr Ser Ser Gln Ser Ser Asn Ser Glu Ser
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aca cct gcc ttc cgc ctg gct gac agc agg act aac cca 1949
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 35 40 45

20 Glu Ser Ala Ala Gly Ser Lys Leu Val Leu Arg Cys Glu Thr Ser
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Ser Glu Tyr Ser Ser Leu Arg Phe Lys Trp Phe Lys Asn Gly Asn
 65 70 75

Glu Leu Asn Arg Lys Asn Lys Pro Gln Asn Ile Lys Ile Gln Lys
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25 Lys Pro Gly Lys Ser Glu Leu Arg Ile Asn Lys Ala Ser Leu Ala
 95 100 105

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30 Asp Ser Ala Ser Ala Asn Ile Thr Ile Val Glu Ser Asn Glu Ile
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Ile Thr Gly Met Pro Ala Ser Thr Glu Gly Ala Tyr Val Ser Ser
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Glu Ser Pro Ile Arg Ile Ser Val Ser Thr Glu Gly Ala Asn Thr
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Lys Cys Ala Glu Lys Glu Lys Thr Phe Cys Val Asn Gly Gly Glu
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	Met Met Asn Ile Ala Asn Gly Pro His His Pro Asn Pro Pro Pro	290	295	300
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	Ser Ser Glu His Ile Val Glu Arg Glu Ala Glu Thr Ser Phe Ser	320	325	330
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	Gln Thr Pro Ser His Ser Trp Ser Asn Gly His Thr Glu Ser Ile	350	355	360
	Leu Ser Glu Ser His Ser Val Ile Val Met Ser Ser Val Glu Asn	365	370	375
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	Gly Thr Gly Gly Pro Arg Glu Cys Asn Ser Phe Leu Arg His Ala	395	400	405
30	Arg Glu Thr Pro Asp Ser Tyr Arg Asp Ser Pro His Ser Glu Arg	410	415	420
	Tyr Val Ser Ala Met Thr Thr Pro Ala Arg Met Ser Pro Val Asp	425	430	435
	Phe His Thr Pro Ser Ser Pro Lys Ser Pro Pro Ser Glu Met Ser	440	445	450
35	Pro Pro Val Ser Ser Met Thr Val Ser Lys Pro Ser Met Ala Val	455	460	465
	Ser Pro Phe Met Glu Glu Glu Arg Pro Leu Leu Leu Val Thr Pro	470	475	480

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	Gln Glu Tyr Glu Pro Ala Gln Glu Pro Val Lys Lys Leu Ala Asn	
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	Arg Leu Glu Val Asp Ser Asn Thr Ser Ser Gln Ser Ser Asn Ser	
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	Glu Ser Glu Thr Glu Asp Glu Arg Val Gly Glu Asp Thr Pro Phe	
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	Ala Phe Arg Leu Ala Asp Ser Arg Thr Asn Pro Ala Gly Arg Phe	
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   Gly Lys Gly Lys Lys Lys Glu Arg Gly Ser Gly Lys Lys
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   ccg gag tcc gcg gcg ggc agc cag agc cca gcc ttg cct 568
   Pro Glu Ser Ala Ala Gly Ser Gln Ser Pro Ala Leu Pro
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10  ccc caa ttg aaa gag atg aaa agc cag gaa tcg gct gca 607
   Pro Gln Leu Lys Glu Met Lys Ser Gln Glu Ser Ala Ala
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   ggt tcc aaa cta gtc ctt cgg tgt gaa acc agt tct gaa 646
   Gly Ser Lys Leu Val Leu Arg Cys Glu Thr Ser Ser Glu
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   tac tcc tct ctc aga ttc aag tgg ttc aag aat ggg aat 685
   Tyr Ser Ser Leu Arg Phe Lys Trp Phe Lys Asn Gly Asn
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20  gaa ttg aat cga aaa aac aaa cca caa aat atc aag ata 724
   Glu Leu Asn Arg Lys Asn Lys Pro Gln Asn Ile Lys Ile
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   caa aaa aag cca ggg aag tca gaa ctt cgc att aac aaa 763
   Gln Lys Lys Pro Gly Lys Ser Glu Leu Arg Ile Asn Lys
       90              95              100

25  gca tca ctg gct gat tct gga gag tat atg tgc aaa gtg 802
   Ala Ser Leu Ala Asp Ser Gly Glu Tyr Met Cys Lys Val
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   atc agc aaa tta gga aat gac agt gcc tct gcc aat atc 841
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   Thr Ile Val Glu Ser Asn Glu Ile Ile Thr Gly Met Pro
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   Ala Ser Thr Glu Gly Ala Tyr Val Ser Ser Glu Ser Pro
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40  tca tct aca tct aca tcc acc act ggg aca agc cat ctt 997
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10 gat cgc tgc caa aac tac gta atg gcc agc ttc tac aag 1153
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	Ser Pro Phe Met Glu Glu Glu Arg Pro Leu Leu Leu Val	
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 40 Ser Glu Tyr Ser Ser Leu Arg Phe Lys Trp Phe Lys Asn Gly Asn
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 Cys Phe Met Val Lys Asp Leu Ser Asn Pro Ser Arg Tyr Leu Cys
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aag aag gag cga ggc tcc ggc aag aag ccg gag tcc gcg 573
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Ala Gly Ser Gln Ser Pro Ala Leu Pro Pro Gln Leu Lys
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15 gag atg aaa agc cag gaa tcg gct gca ggt tcc aaa cta 651
Glu Met Lys Ser Gln Glu Ser Ala Ala Gly Ser Lys Leu
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gtc ctt cgg tgt gaa acc agt tct gaa tac tcc tct ctc 690
20 Val Leu Arg Cys Glu Thr Ser Ser Glu Tyr Ser Ser Leu
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aga ttc aag tgg ttc aag aat ggg aat gaa ttg aat cga 729
Arg Phe Lys Trp Phe Lys Asn Gly Asn Glu Leu Asn Arg
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aaa aac aaa cca caa aat atc aag ata caa aaa aag cca 768
25 Lys Asn Lys Pro Gln Asn Ile Lys Ile Gln Lys Lys Pro
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ggg aag tca gaa ctt cgc att aac aaa gca tca ctg gct 807
Gly Lys Ser Glu Leu Arg Ile Asn Lys Ala Ser Leu Ala
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30 gat tct gga gag tat atg tgc aaa gtg atc agc aaa tta 846
Asp Ser Gly Glu Tyr Met Cys Lys Val Ile Ser Lys Leu
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gga aat gac agt gcc tct gcc aat atc acc atc gtg gaa 885
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tca aac gag atc atc act ggt atg cca gcc tca act gaa 924
Ser Asn Glu Ile Ile Thr Gly Met Pro Ala Ser Thr Glu
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40 Gly Ala Tyr Val Ser Ser Glu Ser Pro Ile Arg Ile Ser
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aca tcc acc act ggg aca agc cat ctt gta aaa tgt gcg 1041
 5 Thr Ser Thr Thr Gly Thr Ser His Leu Val Lys Cys Ala
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gag aag gag aaa act ttc tgt gtg aat gga ggg gag tgc 1080
 Glu Lys Glu Lys Thr Phe Cys Val Asn Gly Gly Glu Cys
 185 190 195

10 ttc atg gtg aaa gac ctt tca aac ccc tcg aga tac ttg 1119
 Phe Met Val Lys Asp Leu Ser Asn Pro Ser Arg Tyr Leu
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tgc aag tgc cca aat gag ttt act ggt gat cgc tgc caa 1158
 15 Cys Lys Cys Pro Asn Glu Phe Thr Gly Asp Arg Cys Gln
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aac tac gta atg gcc agc ttc tac agt acg tcc act ccc 1197
 Asn Tyr Val Met Ala Ser Phe Tyr Ser Thr Ser Thr Pro
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20 ttt ctg tct ctg cct gaa tagga gcatgctcag ttggtgctgc 1240
 Phe Leu Ser Leu Pro Glu
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 35 <213> Homo sapiens

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25	Cys	Phe	Met	Val	Lys	Asp	Leu	Ser	Asn	Pro	Ser	Arg	Tyr	Leu	Cys	
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	Met	Cys	Val	Val	Ala	Tyr	Cys	Lys	Thr	Lys	Lys	Gln	Arg	Lys	Lys	
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35	Leu	His	Asp	Arg	Leu	Arg	Gln	Ser	Leu	Arg	Ser	Glu	Arg	Asn	Asn	
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Glu Asn Val Gln Leu Val Asn Gln Tyr Val Ser Lys Asn Val Ile
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 5 Thr Ser His Tyr Thr Ser Thr Ala His His Ser Thr Thr Val Thr
 335 340 345
 Gln Thr Pro Ser His Ser Trp Ser Asn Gly His Thr Glu Ser Ile
 350 355 360
 10 Leu Ser Glu Ser His Ser Val Ile Val Met Ser Ser Val Glu Asn
 365 370 375
 Ser Arg His Ser Ser Pro Thr Gly Gly Pro Arg Gly Arg Leu Asn
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 cctgcgccga gagccgtccg cgtagagcgc tccgtctccg gcgag atg 498
 Met
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tcc gag cgc aaa gaa ggc aga ggc aaa ggg aag ggc aag 537
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aag aag gag cga ggc tcc ggc aag aag ccg gag tcc gcg 576
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gcg ggc agc cag agc cca gcc ttg cct ccc caa ttg aaa 615
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gag atg aaa agc cag gaa tcg gct gca ggt tcc aaa cta 654
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aaa aac aaa cca caa aat atc aag ata caa aaa aag cca 771
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 80 85 90

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 20 Gly Lys Ser Glu Leu Arg Ile Asn Lys Ala Ser Leu Ala
 95 100 105

gat tct gga gag tat atg tgc aaa gtg atc agc aaa tta 849
 Asp Ser Gly Glu Tyr Met Cys Lys Val Ile Ser Lys Leu
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gga aat gac agt gcc tct gcc aat atc acc atc gtg gaa 888
 25 Gly Asn Asp Ser Ala Ser Ala Asn Ile Thr Ile Val Glu
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tca aac gag atc atc act ggt atg cca gcc tca act gaa 927
 30 Ser Asn Glu Ile Ile Thr Gly Met Pro Ala Ser Thr Glu
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 Gly Ala Tyr Val Ser Ser Glu Ser Pro Ile Arg Ile Ser
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5	tgc aag tgt cca aat gag ttt act ggt gat cgc tgc caa	1161
	Cys Lys Cys Pro Asn Glu Phe Thr Gly Asp Arg Cys Gln	
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10	tac cag aag aga gtg ctg acc ata acc ggc atc tgc atc	1239
	Tyr Gln Lys Arg Val Leu Thr Ile Thr Gly Ile Cys Ile	
	240 245	
	gcc ctc ctt gtg gtc ggc atc atg tgt gtg gtg gcc tac	1278
	Ala Leu Leu Val Val Gly Ile Met Cys Val Val Ala Tyr	
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	tgc aaa acc aag aaa cag cgg aaa aag ctg cat gac cgt	1317
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20	Leu Arg Gln Ser Leu Arg Ser Glu Arg Asn Asn Met Met	
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	Glu Asn Val Gln Leu Val Asn Gln Tyr Val Ser Lys Asn	
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	Val Ile Ser Ser Glu His Ile Val Glu Arg Glu Ala Glu	
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	Thr Ser Phe Ser Thr Ser His Tyr Thr Ser Thr Ala His	
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	cac tcc act act gtc acc cag act cct agc cac agc tgg	1551
35	His Ser Thr Thr Val Thr Gln Thr Pro Ser His Ser Trp	
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	agc aac gga cac act gaa agc atc ctt tcc gaa agc cac	1590
	Ser Asn Gly His Thr Glu Ser Ile Leu Ser Glu Ser His	
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	Ser Val Ile Val Met Ser Ser Val Glu Asn Ser Arg His	
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    Ser Ser Pro Thr Gly Gly Pro Arg Gly Arg Leu Asn Gly
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    gcc aga gaa acc cct gat tcc tac cga gac tct cct cat 1746
    Ala Arg Glu Thr Pro Asp Ser Tyr Arg Asp Ser Pro His
      405                      410                      415

10  agt gaa agg taaaa ccgaaggcaa agctactgca gaggagaaac 1790
    Ser Glu Arg
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    ctttgatgcg gaaggtgcag cacatggagt ttccagctct ggccatgggc 2040

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	Pro	His	Gly	Thr	Leu	Tyr	Arg	Thr	Asp	Ile	Gly	Leu	Pro	His	Cys	95	100	105
10	Gly	Tyr	Ser	Met	Gly	Ala	Gly	Ser	Asp	Ala	Asp	Met	Glu	Ala	Asp	110	115	120
	Thr	Val	Leu	Ser	Pro	Glu	His	Pro	Val	Arg	Leu	Trp	Gly	Arg	Ser	125	130	135
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	Pro	Ser	Pro	Ala	Pro	Thr	Asp	His	Ser	Leu	Ser	Gly	Glu	Pro	Pro	215	220	225
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	Arg	Pro	Leu	Pro	Arg	Ser	Thr	Phe	Ala	Arg	Pro	Ala	Phe	Asn	Leu	320	325	330

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 15 Gly Glu Cys Phe Met Val Lys Asp Leu Ser Asn Pro Ser Arg Tyr
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 35 Met Asp Val
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